



OPEN ACCESS

EDITED BY

Vinay Kumar,
The Pennsylvania State University,
United States

REVIEWED BY

Lorenzo Da Dalt,
University of Milan, Italy
Suman Asalla,
The Ohio State University, United States

*CORRESPONDENCE

Ming-Sheng Zhou

✉ zhoums@symc.edu.cn

Lu Zhang

✉ zhanglu0412@symc.edu.cn

[†]These authors have contributed equally to this work

RECEIVED 19 July 2024

ACCEPTED 03 September 2024

PUBLISHED 20 September 2024

CITATION

Kang M, Jia H, Feng M, Ren H, Gao J, Liu Y, Zhang L and Zhou M-S (2024) Cardiac macrophages in maintaining heart homeostasis and regulating ventricular remodeling of heart diseases. *Front. Immunol.* 15:1467089. doi: 10.3389/fimmu.2024.1467089

COPYRIGHT

© 2024 Kang, Jia, Feng, Ren, Gao, Liu, Zhang and Zhou. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](#). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Cardiac macrophages in maintaining heart homeostasis and regulating ventricular remodeling of heart diseases

Mengjie Kang^{1†}, Hui Jia^{1,2†}, Mei Feng¹, Haolin Ren³, Junjia Gao⁴, Yueyang Liu^{1,5}, Lu Zhang^{1*} and Ming-Sheng Zhou^{1*}

¹Science and Experiment Research Center, Shenyang Medical College & Shenyang Key Laboratory of Vascular Biology, Science and Experimental Research Center, Shenyang Medical College, Shenyang, China, ²School of Traditional Chinese Medicine, Shenyang Medical College, Shenyang, China, ³Department of Radiology, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, ⁴Department of Cardiology, Second Affiliated Hospital, Shenyang Medical College, Shenyang, China, ⁵School of Pharmacy, Shenyang Medical College, Shenyang, China

Macrophages are most important immune cell population in the heart. Cardiac macrophages have broad-spectrum and heterogeneity, with two extreme polarization phenotypes: M1 pro-inflammatory macrophages (CCR2⁻ly6C^{hi}) and M2 anti-inflammatory macrophages (CCR2⁻ly6C^{lo}). Cardiac macrophages can reshape their polarization states or phenotypes to adapt to their surrounding microenvironment by altering metabolic reprogramming. The phenotypes and polarization states of cardiac macrophages can be defined by specific signature markers on the cell surface, including tumor necrosis factor α , interleukin (IL)-1 β , inducible nitric oxide synthase (iNOS), C-C chemokine receptor type (CCR)2, IL-4 and arginase (Arg)1, among them, CCR2^{+/+} is one of most important markers which is used to distinguish between resident and non-resident cardiac macrophage as well as macrophage polarization states. Dedicated balance between M1 and M2 cardiac macrophages are crucial for maintaining heart development and cardiac functional and electric homeostasis, and imbalance between macrophage phenotypes may result in heart ventricular remodeling and various heart diseases. The therapy aiming at specific target on macrophage phenotype is a promising strategy for treatment of heart diseases. In this article, we comprehensively review cardiac macrophage phenotype, metabolic reprogramming, and their role in maintaining heart health and mediating ventricular remodeling and potential therapeutic strategy in heart diseases.

KEYWORDS

macrophages, heart diseases, metabolic reprogramming, polarization, cardiac remodeling

1 Introduction

Macrophages are an important type of immune effector cells, which inhabit almost all tissues and organs of the body, including the heart. Macrophages play important roles not only in tissue immunity and inflammation, but also in organ development, homeostasis and remodeling (1–3). The broad-spectrum of macrophages depends on their heterogeneity and plasticity, and macrophages can modify their phenotype or polarization, adapting to their microenvironment (4). The phenotype of macrophages manifests as two extreme polarizations: Classically activated M1 pro-inflammatory macrophages, also known as pro-inflammatory M1 macrophage, and alternatively activated M2 anti-inflammatory macrophages. M1 macrophages are involved in the initiation and maintenance of inflammation, recruiting other immune cells into inflamed tissue, and releasing inflammatory cytokines to kill foreign pathogens or microorganisms. M2 macrophages promote the resolution of inflammation, engulf apoptotic cells, drive collagen deposition, coordinate tissue integrity, and release anti-inflammatory mediators (5, 6). The delicate balance between M1 and M2 macrophages is crucial for maintenance of homeostasis. The imbalance of the two subtypes of macrophage may lead to many diseases including cardiovascular diseases, metabolic diseases and cancer (1).

Increasing evidence has shown that cardiac resident macrophages (CRMs) importantly contribute to the maintenance of heart health and the development of heart diseases (7). The origin of CRMs can be from the yolk sacs during embryonic development or hematopoietic cells-derived monocytes (8). Embryonic CRMs exist from embryonic development and persist into adulthood, and are mainly involved in heart development, homeostasis and repair (9). Upon the stimulation of inflammation or in the situation of heart diseases, the monocytes-derived macrophages are recruited, and CRMs are expanded in the heart to combat the injury stimuli and induce heart remodeling (10). Similar to macrophages residing in other tissues, CRMs can change their phenotype or polarization and undergo metabolic reprogramming in response to physiological or pathophysiological alterations in their microenvironment (11). Polarized macrophages exhibit metabolic changes, such as glycolysis, oxidative phosphorylation (OXPHOS), fatty acid oxidation (FAO), and amino acid metabolism. In addition to infectious diseases, macrophage polarization and metabolic programming also participate in the pathogenesis of cardiovascular diseases, especially heart ventricular remodeling diseases (12). This article focuses on reviewing the progress on the CRMs in maintaining heart health, mediating heart ventricular remodeling-related diseases and potential therapeutic strategy for heart diseases.

2 Macrophage polarization and metabolic reprogramming

Macrophages hold extreme plasticity and heterogeneity, and they can quickly adapt to the environment through modifying their

phenotype or polarization (13). Unstimulated macrophages are under a quiescent or non-polarized state, which is also called M0 macrophage. Upon immune challenge, M0 macrophages have the ability to polarize into two distinct cell phenotypes of M1 inflammatory macrophages or M2 macrophages with the possibility to switch from M2 to M1 macrophage. M1 macrophages are typically activated by lipopolysaccharides (LPS), tumor necrosis factor (TNF) α or interferon (IFN) γ (14). M1 macrophages support a glycolytic activity to rapidly produce adenosine triphosphate (ATP) and efficiently fuel the cell at early stage of infection (15). The metabolic reprogramming toward aerobic glycolysis is called the “Warburg effect”, which is similar to metabolic reprogramming in cancer cells (16). Activated M1 macrophages produce large amounts of inflammatory cytokines and reactive oxygen species (ROS), which are beneficial for killing invaded microorganisms and controlling inflammation (17). However, the released inflammatory cytokines and ROS from M1 macrophages may also cause tissue chronic inflammation and injury, which is often present in cardiovascular diseases. M2 macrophages are usually induced by IL-4, IL-13 or transforming growth factor (TGF)- β (1). M2 macrophages have an intact tricarboxylic acid (TCA) cycle and use mitochondrial OXPHOS and FAO to generate ATP (10, 18, 19). M2 macrophages are involved in clearing apoptotic cells, driving collagen deposition and angiogenesis, and releasing anti-inflammatory mediators for anti-inflammatory and promoting tissue repair process (20). In addition, macrophages may have immune memory function on pathogen stimulation. M1/M2 macrophage polarization is always linked to the metabolic reprogramming of glucose, FAO, and amino acid metabolism (Figure 1).

2.1 M1 macrophage polarization and metabolism

Macrophages have surface and cytosolic sensors, such as toll-like receptors (TLRs) on cell surface and NOD-like receptors (NLRs) in the cytoplasm, which can respond to foreign pathogens or microbial products to trigger immune responses (21, 22). For example, LPS binds to TLRs on the macrophage surface, which subsequently activates downstream molecules to induce the activation of nuclear factor (NF)- κ B pathway and release inflammatory cytokines, such as IL-1 β , IL-6, chemokine ligands (CXCL)9, CXCL10 and monocyte chemoattractant protein (MCP)-1 (23, 24). Some pathogens may be engulfed by macrophages and bind to NLRs in the cytoplasm to form inflammasomes, which activate NF- κ B inflammatory pathway and promote macrophage activation (22). Interestingly, the stimulation of these sensors not only induce immune response but also drive macrophage into a Warburg-like response (25, 26). LPS-induced Warburg shift is a metabolic reprogramming that shares many similarities but differs significantly from the Warburg response in cancer cells. Cancer cells use glucose-6-phosphate as a fuel for lipid and nucleotide synthesis, and metabolize pyruvate to support amino acid and fatty acid synthesis for cell growth, while inflammatory responses

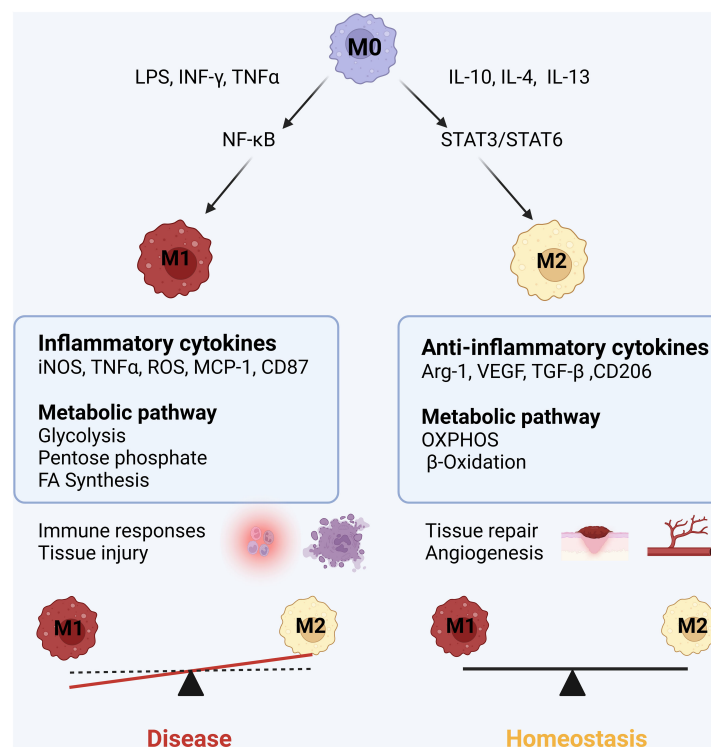


FIGURE 1

M1 and M2 macrophage polarization: Upon inflammatory (LPS, INF γ , TNF α) stimuli, macrophages are polarized into inflammatory M1 phenotype by activating NF- κ B inflammatory pathway, simultaneously, the metabolic pathway of M1 macrophages shifts into glycolysis and pentose phosphate pathway. M1 macrophages release inflammatory mediators including inducible nitric oxide synthase (iNOS), reactive oxygen species (ROS), reactive nitric species (RNS), tumor necrotic factor (TNF) α , interleukin (IL)-1 β , IL-6, monocyte chemokine protein (MCP)-1, leading to cytotoxicity and tissue injury. When inflammatory stimuli subsides or anti-inflammatory cytokines IL-10, IL-4 and IL-13 increase, macrophages are polarized into M2 anti-inflammatory macrophage associated with the switch of metabolic pathway into oxidative phosphorylation (OXPHOS) and fatty acid β -oxidation. M2 macrophages release arginase 1 (Arg1), VEGF, IL-10 and TGF- β 1, promoting to tissue repair and wound healing.

mediated by TLRs produce the metabolites related to the antimicrobial activities, such as lactate, iNOS and ROS (26, 27).

Pro-inflammatory M1 macrophages shift their metabolism to aerobic glycolysis to meet the increasing energy requirement, and this metabolic adjustment is similar to the Warburg effect, referring to the metabolic reprogramming of macrophages, where macrophages rely solely on glycolysis to obtain energy, and the TCA cycle is inactivated even in the presence of oxygen (28). Glucose is metabolized through glycolysis to produce pyruvate and two molecules of ATP. Although glycolysis is relatively inefficient in ATP generation, the rate of glycolysis is very fast, the rapid increase in glycolytic metabolism is crucial for maintaining macrophage phagocytic activity (29). More importantly, increased aerobic glycolysis may compensate for the decreased ATP synthesis due to inflammation inhibition of mitochondrial respiration, which may help to maintain cellular ATP level and mitochondrial membrane potential (30).

Glycolysis can provide metabolic intermediates for the synthesis of fatty acids, amino acids and ribose, which are necessary for cellular metabolic adaptation. In addition, during the glycolytic oxidative stage, NADP $^+$ is converted into NADPH. NADPH can be utilized by NADPH oxidase to produce ROS, which activates redox

sensitive transcriptional factor NF- κ B, promoting macrophage activation (31). In many circumstances, the activation of M1 macrophages is associated with hypoxic microenvironments, and hypoxia can profoundly affect macrophage function. Hypoxia inducible factor (HIF) 1 α is an important transcriptional factor, which regulates a large amount of gene expressions, including the genes related to glycolysis and inflammatory mediators (32). For example, HIF1 α upregulates glucose transporter (GLUT)1 to facilitate rapid uptake of glucose and lactate dehydrogenase to promote the conversion of pyruvate into lactate, which are crucial for metabolic glycolysis adaption of macrophage (33). HIF1 α also upregulates pyruvate dehydrogenase kinase which inactivates pyruvate dehydrogenase to limit pyruvate into Krebs cycle. Therefore, HIF1 α is an important regulator factor that promotes macrophage metabolic reprogramming and aerobic glycolysis (24, 34). The metabolic reprogramming of M1 macrophages mainly manifests by shutting down TCA cycle and shifting it into aerobic glycolysis, which helps M1 macrophages to maintain their inflammatory phenotype and produce pro-inflammatory cytokines and chemokines (Figure 2).

In glucose metabolism, two key enzymes are involved in regulating glycolysis and macrophage M1 polarization:

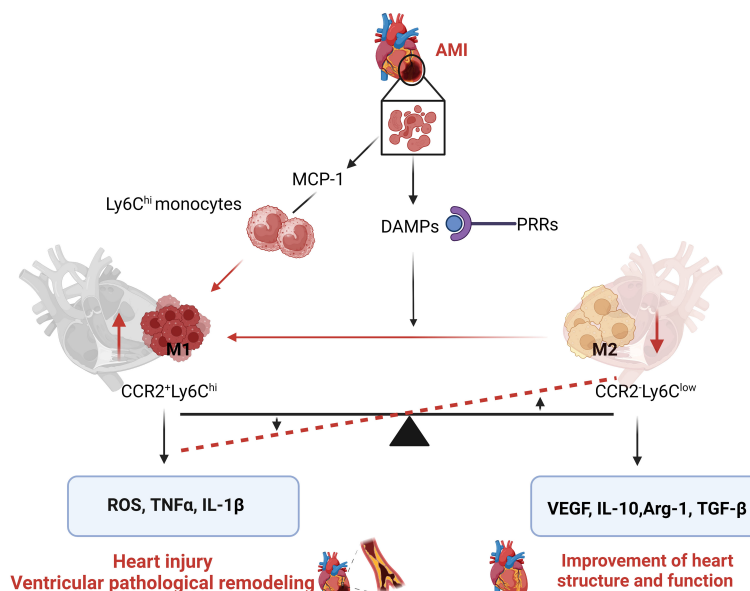


FIGURE 2

Metabolic reprogramming of M1/M2 macrophages: M1 inflammatory macrophages activate HIF1 α pathway, which upregulates two key enzymes of glycolysis: 6-phosphofructose-2-kinase B (PFKFB) and pyruvate kinase M2 (PKM2), and increases activity of lactate dehydrogenase (LDH) that promotes the conversion of pyruvate to lactic acid, thus, increasing glycolysis. ATP generation from rapid glycolysis in turn inhibits mitochondria respiratory chain to reduce ATP generation from tricarboxylic acid cycle (TCA) through reduced key enzymes of TCA cycle, such as isocitrate dehydrogenase (IDH) and succinate dehydrogenase (SDH). The decreased activity of IDH and SDH increases the accumulation of substrates isocitrate and succinate in mitochondria which leak into the cytoplasm to continuously activate NF- κ B and HIF1 α , thus maintaining M1 macrophage activation and glycolysis. In other way, M2 macrophages have low level of glycolysis with intact TCA cycle, they generate ATP through mitochondria OXPHOS. In addition, M2 macrophages express high level of Arg1, which inhibit iNOS-derived NO production, which promotes the entry of pyruvate into the TCA cycle and inhibits glycolysis.

6-phosphofructose-2-kinase/fructose-2,6-bisphosphatase (PFKFB) and pyruvate kinase M (PKM)2, which are highly expressed in activated immune cells (35, 36). M1 macrophages mainly express PFKFB3 subtype and pyruvate kinase subtype 2. The PFKFB3 has a relatively low efficiency in catalyzing the conversion of 2,6-diphosphate fructose to 6-phosphate fructose; thereby increasing the glycolytic flux (37). PKM2 plays multiple roles in macrophage metabolism and polarization. In fact, when highly expressed, PKM2 exists in a balance between monomeric or dimeric enzymes without enzymatic activity and tetramers with enzymatic activity. Inactive enzymes of PKM2 monomer and dimer transfer to the nucleus and affect glycolysis by activating the expression of HIF1 α gene, while tetramers with enzyme activity retains in the cytoplasm to promote glycolysis and M1 macrophage polarization (38).

It is well known that M1 macrophages are involved in the pathogens of cardiovascular diseases, especially atherosclerosis (1, 39). Atherosclerosis is considered as a chronic inflammatory vascular disease, and macrophages play a key role in the progress of atherosclerosis. Cluster of differentiation (CD)36 is an important scavenger receptor on the surface of macrophages, involved in the uptake of oxidized low-density lipoprotein (oxLDL), and oxLDL induces macrophage inflammation and activation (40). We have shown that nicotine can upregulate CD36 expression in macrophage which potentiates oxLDL-induced macrophage activation and foam cell formation (41).

2.2 M2 macrophage polarization and metabolism

M2 macrophages are induced by crucial cytokines, such as IL-4, IL-10, IL-13 and TGF- β , which are secreted by Th-2 immune cells (42). IL-4 and IL-13 induce M2 macrophage polarization by activating signal transducer and activator transcription (STAT)6 signaling, and IL-10 activates M2 macrophages by activating STAT3 (43, 44). M2 macrophages express high levels of CD206, CD163 and Arg1, and produce profibrotic factors TGF- β , vascular endothelium growth factor (VEGF) and insulin-like growth factor, therefore actively contributing to the resolution of inflammation, angiogenesis and tissue repair and healing. TGF- β , Arg1 and VEGF increase the production of collagen and angiogenesis, which favors tissue remodeling and wound repair (45). M2 macrophages have anti-inflammatory properties. In many cardiovascular diseases, activating M2 macrophages is considered to have protective effects. For example, it has been shown that activating M2 macrophages can increase the stability and regression of atherosclerotic plaques (46).

M2 macrophages fuel glucose and fatty acid, and rely on OXPHOS and FAO to generate ATP. Despite the controversy, it is generally believed that M2 macrophages are rarely dependent on glycolysis (12, 47). The mechanisms by which M2 macrophages exhibit low glycolytic activity are not fully understood. The

following mechanisms may be related to low level glycolysis of M2 macrophages: Firstly, M2 macrophages have intact TCA cycle. They can provide ATP through mitochondrial OXPHOS (28). Next, PFKFB1 is a regulator of glycolysis, M2 macrophages have selective expression of PFKFB1, which can efficiently catabolize glycolytic activator fructose 2,6-bisphosphate to 6-phosphate, therefore limiting the glycolytic rate (29). Finally, M2 macrophages overexpress Arg1. As both Arg1 and iNOS use arginine as substrate, Arg1 may compete with iNOS for arginine substrate; thereby blocking iNOS-mediated NO production, which may reduce glycolysis (48) (Figure 2).

3 Cardiac macrophages in the maintenance of heart homeostasis

3.1 Cardiac macrophage lineage

The simplified definition of M1/M2 macrophages may fit in the situation of the *in vitro* experiments, and cannot accurately describe the characteristic of the cardiac macrophage phenotypes. Cardiac macrophages *in vivo* are more complex and heterogenous (49). Cardiac macrophage phenotype may be continuous in physiological and pathological states rather than just two types of polarized cells. CRMs have specific phenotypes and function in different cardiac regions and stages (50). According to the expressions of CCR2 and human leukocyte antigen DR (HLA-DR), human cardiac macrophages are classified into three different subgroups of CCR2⁺HLA-DR^{lo}, CCR2⁺HLA-DR^{hi}, CCR2⁻HLA-DR^{hi}, and HLA-DR is a human homolog of the major histocompatibility complex (MHC)-II (51). According to the expression of the lymphocyte antigen 6 complex locus C (Ly6C) and MHC-II, mouse CRMs can distinguish four different phenotypes: CX3CR1⁺CCR2⁻Ly6C⁻MHC-II⁻ (MP1), CX3CR1⁺CCR2⁻Ly6C⁻MHC-II⁺ (MP2), CX3CR1⁺CCR2⁺Ly6C⁺MHC-II⁻ (MP3) and CX3CR1⁺CCR2⁺Ly6C⁺MHC-II⁺ (MP4) macrophages (52). Based on the expressions of the biomarkers including lymphatic endothelial hyaluronic acid receptor (LYVE)1, phosphatidylserine receptor T cell immunoglobulin and mucin containing domain (TIMD)4, folate receptor β (FOLR2, TLF), CCR2, and MHC-II, Dick et al. (53) propose a more complex method of CRMs classification. They classify CRMs into three subgroups: 1) TLF⁺ CRMs expressing CX3CR1^{lo}TIMD4⁺LYVE1⁺TLF⁺MHC-II^{lo}CCR2⁻; 2) CCR2⁺ CRMs expressing CX3CR1^{hi}CCR2⁺CD11c⁺MHC-II^{hi}TIMD4⁻LYVE1⁻TLF⁻; 3) MHC-II^{hi} CRMs expressing CX3CR1^{hi}MHC-II^{hi}TIMD4⁻LYVE1⁻TLF⁻CCR2⁻. Among the group, CCR2⁺ CRMs have limited self-renewal ability and are constantly replaced by the circulating monocytes, TLF⁺ CRMs is almost entirely maintained by self-proliferation *in situ*, and MHC-II^{hi} CRMs only have a few subpopulations updated by monocytes. CCR2^{+/+} is considered as a key biomarker to distinguish between resident and non-resident CRMs which are conserved in humans, rats and mice, and TLF⁺CCR2⁻ and MHC-II^{hi} CRMs are considered CCR2⁻ CRMs (54).

With the development of novel biotechnologies, such as genetic fate mapping and lineage tracing, scientists can trace the origin of

CRMs (9). It is generally believed that CRMs have two independent origins: One type of CRMs is established during the embryonic stage and embryonic development and can be self-sustaining and regenerating, this type of CRMs expresses CCR2⁻ and originates from yolk sacs, another type of CRMs expresses CCR2⁺ and originates from circulating monocytes, CCR2⁺ CRMs are continuously replaced by the circulating monocytes (55).

3.2 Macrophage regulation of heart electric activity and homeostasis

The generation and conduction of action potentials is an important feature of the heart. Under normal circumstances, the sinoatrial node generates action potential and controls heart electric activity, which propagate to all myocardial cells throughout the heart conduct system, inducing heart contraction (56). The heart contracts in the form of cardiomyocyte syncytium, and the synchronous contraction of the heart chambers is crucial for maintaining cardiac ejection function, and the completion of this function mainly depends on the high coordination of electric activity and contractile proteins in myocardial cells (57). It is well known that the conduction of cardiac electric impulses between myocardial cells is through gap junction (58). The simultaneous activation of action potentials in myocardial cells and an organized heart contraction require a normal gap junction communication (59, 60). Gap junctions are non-selective pores on myocardial cells, also known as gap junction channels, which allow ions and other small molecules to pass freely between myocardial cells, providing rapid conduction of electric impulse and electrical continuity between myocardial cells (61, 62). A single myocardial cell membrane only has gap junction hemichannels. These hemichannels converge at intercalated discs on myocardial cell membrane, which bind end to end with the hemichannels from adjacent myocardial cells, forming an intact gap junction channel (63).

The heart has four types of connexin proteins, including connexin (Cx)43, Cx45, Cx 40 and Cx30.2 (60, 64). Among them, Cx43 is the most abundant connexin protein, and is almost expressed in all parts of the heart except for the sinoatrial node. Cardiac gap junction intercellular communication is mainly controlled by post-translational regulation of Cx43. There are 21 phosphorylation sites at the C-terminal of Cx43, which can be phosphorylated or dephosphorylated by many protein kinases or phosphatases, thereby changing the localization of Cx43, the ion transport and electrical conductivity in gap junction (61, 65).

As mentioned above, CRMs are widely distributed in the heart. It has been shown that CRMs, which mainly express CCR2⁻, are also present in cardiac autorhythmic tissues, including sinoatrial node, atrioventricular node (AVN) and His-Bundle (66). Recently, Hulsmans et al. (67) demonstrated that CRMs, which were enriched in mouse and human AVN, had physical contact with neighboring conductive myocardial cells and facilitated electrical conduction in distal AVN and AV His-bundle through Cx43-containing gap junction. Using green fluorescent protein (GFP) labeled macrophages, they demonstrated that these macrophages are longitudinally distributed along the AV His-Bundle, with

cytoplasmic portions extending and reaching long distance cardiac myocytes. Cx43-containing gap junction provides a structural framework for the interaction and communication between CRMs and conductive cardiac myocytes (68). Cell co-culture studies reveal a bidirectional regulation between macrophages and cardiac myocytes, and the depolarization of cardio myocytes induce the depolarization of macrophages, as only macrophages coupled with cardio myocytes generate depolarization potential (69). On the other hand, the electrical activity of macrophages, in turn, significantly affects the action potential of cardio myocytes, including reduction in the negative amplitude of resting potential, shortening action potential duration and refractory period, thus accelerating the electric conduction of cardio myocyte (68, 70). Cx43 plays an important role in electrical interaction and communication between CRMs and conductive myocardial cells, as the deletion of Cx43 in macrophages can eliminate the electrical interaction and communication between macrophage and myocardial cells (71). Furthermore, the conditional depletion of macrophage Cx43 or macrophage ablation can develop severe AV block in mice. These studies suggest that CRMs are required for the maintaining normal heart conduction in the AVN (67).

Although the mechanism by which macrophages participate in myocardial electrophysiology is not understood, Cx43 may be a significant medium that connects macrophages and conductive myocardial cells to regulate cardiac electrical activity. It has been shown that post-transcription regulation of Cx43 phosphorylation can alter gap junction function, affecting cardiac conduction (72). It has been shown that M1 polarization macrophage can upregulate potassium channel KCa3.1, which promotes Ca^{2+} influx in myocardial cells through Cx43, resulting in prolonged action potential durations (APD) (73, 74). In addition, the pro-inflammatory cytokines released from activated macrophage can affect cardiac electrical activity and induce cardiac electrical remodeling (75, 76). Thus, it tempts to hypothesize that CRMs may affect gap junction and cardiac electrical activity in myocardial cells through post-modification of Cx43 phosphorylation.

3.3 Macrophages maintain cardiac homeostasis

Macrophages are the most abundant and important immune cells in the heart, and play a critical role in heart development, maintenance of physiological homeostasis and induction of pathological ventricular remodeling in the heart diseases (9). Like macrophages in other tissues, CRMs have heterogeneity and functional plasticity to adapt to their microenvironment and meet the needs of their home tissue (12). CCR2⁺ CRMs are the main population, and expanded through self-renewal and proliferation *in situ* in the health heart, upon inflammatory or injury stimuli, the circulating monocytes can migrate into cardiac tissue and differentiate into cardiac macrophages, thus cardiac macrophages can be rapidly replenished by migrating peripheral monocytes to deal with the changed environment (7). These cardiac macrophages express CCR2⁺ and have strong phagocytic activity.

One mechanism by which cardiac macrophages maintain cardiac homeostasis may be through efferocytosis (77). Efferocytosis is a process by which macrophages recognize and engulf the dead cells, tissue debris or metabolic wastes by phagocytic receptor alone or in combination with other receptors, and is a characteristic function of macrophages. It is well known that myocardial cells are a type of long-lived and rarely self-renewal cells, myocardial cells have high metabolisms and are long-term exposed to intense mechanic stress, which may make cardiomyocytes produce lot of dysfunctional mitochondria and other cargos and eject them into the extracellular compartment (78). CRMs actively eat up these metabolic wastes to maintain cardiac homeostasis by efferocytosis. The depletion of CRMs may result in the accumulation of metabolic wastes and impair heart function.

CRMs maintenance of cardiac homeostasis also exhibit spatiotemporal coordination between CRMs heterogenous and local myocardial cell function (7, 9). It has been proposed that each heart chamber may have different CRM subsets with a specific phenotype and function. For example, the macrophages in the right atrium contain a distinct subset of CRMs that is much smaller than the CRMs in the ventricle tissue and almost non-existent in other chambers (79). In addition, CRMs between the atrium and the ventricle are also significantly different, which may be related to the differences in physiological functions, such as electrophysiological activity and contraction characteristics between atrial and ventricular muscle cells (80). These differences in function and structure between the atrium and ventricle may shape the heterogeneous CRMs subsets to adapt to their physiological function and local microenvironment.

4 The role of CRMs in ventricular remodeling in heart diseases

4.1 Myocardial infarction-induced ventricular remodeling

Myocardial infarction (MI) is a clinical syndrome of acute myocardial ischemic injury caused by partial or complete occlusion of the coronary artery (81). Persistent ischemia/hypoxia induces a series of alterations, including endoplasmic reticulum stress, ROS production, mitochondrial dysfunction, inflammation, cell death and ventricular fibrosis and remodeling. Postinfarction ventricular remodeling is the most prominent pathological alteration, which not only occurs in the region of infarcted myocardial tissue, but also reshapes whole ventricular structure and result in the progressive expansion of ventricle, and the development of HF and ventricular aneurysm (82, 83).

After MI, the heart initiates a serious response to repair damaged heart, and typically undergoes three phases: Inflammation, resolution of inflammation, and ventricular remodeling. Monocytes/macrophages are involved in all three phases and play a decisive role in these processes (84). In the early stage of MI, damaged cardiomyocytes release chemokines,

such as MCP-1, C-C motif ligand (CCL) 2, CCL3. MCP-1 is an important chemokine that attracts bone marrow and spleen-derived Ly6C^{hi} monocytes to cross the vascular wall and migrate into the inflamed tissue, where Ly6C^{hi} monocytes differentiate into M1 inflammatory macrophage and gradually replace native CRMs (85). In addition, damaged cardiomyocytes can release danger-associated molecular proteins (DAMPs), which bind to pattern recognition receptors (PRRs) to form inflammasome to induce the activation of macrophages through activating the NF- κ B pathways (86). Activated M1 macrophages have potent phagocytic activity, which clear up the dead cells, damaged organelles and cellular debris, and help to maintain heart homeostasis and function (1). However, M1 macrophages can also secrete pro-inflammatory cytokines, such as IL-1 β , IL-6, TNF α and increase ROS generation, which further aggravate myocardial cell injury and death, and worsen the microenvironment of myocardial cells, such as exacerbating hypoxia (5). Hypoxia activates the HIF1 α pathway to induce the metabolic reprogramming of macrophages primarily through glycolysis (34, 38). If myocardial ischemia cannot be relieved, continuous inflammation and activated M1 macrophage are the main pathological mechanisms leading to the poor prognosis of MI (87). Approximately 4 days after MI, as inflammation subsides and inflammatory cytokine stimulation

signals weaken, partial M1 inflammatory macrophages in the infarct area begin to transform into anti-inflammatory M2 macrophages (Ly6C^{lo}) with a metabolic reprogramming of OXPHOS (88). M2 macrophages release cytokines, such as IL-10, vascular endothelial growth factor (VEGF), TGF- β , etc., which can inhibit inflammation, promote angiogenesis, collagen and extracellular deposition, and facilitate the reconstruction of heart structure and function (12). However, in most cases, myocardial ischemia at the infarcted area is not relieved, and the damaged and necrotic myocardial tissue serves as an inflammatory signal that continuously stimulates M1 macrophages to release inflammatory cytokines, resulting in an excessive inflammatory response, facilitating the adverse remodeling of myocardium (89). In addition, MI may impair macrophage efferocytosis to limit macrophage capability to clear up cellular debris and damaged organelle (77, 90) (Figure 3).

There are several molecules that can affect the polarization and metabolic reprogramming of macrophages, therefore affecting cardiac remodeling after MI. Ndufs4 encoded in mitochondrial complex I protein is involved in regulating cellular ATP synthesis. Cai et al. (91) have reported that specific KO of Ndufs4 in myeloid cells increase the pro-inflammatory cytokine expression of M1 macrophages, and delay the repair period after MI. In addition,

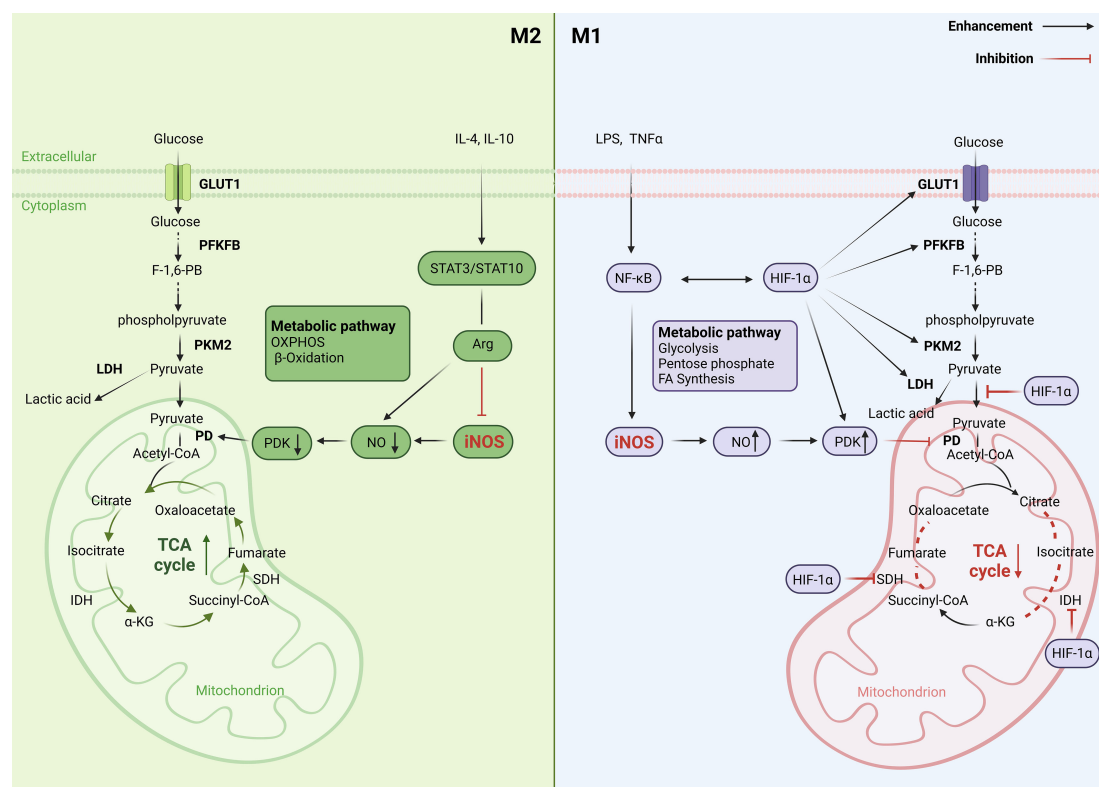


FIGURE 3

Acute myocardial infarction (AMI) alters cardiac macrophage subpopulation to induce pathological cardiac remodeling. After AMI, damaged or dead cardiomyocytes release MCP-1, which attracts circulatory Ly6C^{hi} monocytes to recruit to the infarcted area and differentiate into $\text{CCR2}^+\text{Ly6C}^{\text{hi}}$ macrophage. In other way, danger-associated molecular proteins (DAMPs) released from injury cells bind to pattern recognition receptors (PRRs) and promote the switch of $\text{CCR2}^-\text{Ly6C}^{\text{low}}$ cardiac resident macrophages (CRMs) into CCR2^+ CRMs, resulting in the imbalance between CCR2^+ CRMs and CCR2^- CRMs. CCR2^+ CRMs secrete pro-inflammatory cytokines ROS generation to aggravate myocardial cell injury and death and induce pathological cardiac remodeling.

the efferocytosis of macrophages in KO mice is impaired with the limitation of the proliferation and activation of fibroblasts in the infarction area, leading to adverse repair and ventricular remodeling. Leucine-rich repeating G-protein-coupled receptors (Lgr)4, also known as GPR48, is a member of the Lgr family that is gaining attention as a regulator of macrophage-related immune responses (92). Huang et al. (93) have shown that Lgr4 is highly expressed in activated macrophages, and specific knockout of Lgr4 in myeloid cells significantly improved cardiac function and cardiac remodeling in MI mice associated with reduction in pro-inflammatory Ly6C^{hi} mononuclear/macrophage in the infarct area.

Anti-inflammatory M2 macrophages play a crucial role in limiting adverse remodeling after MI. Zhang et al. (94) revealed that nucleophosmin (NPM)1 is a key regulatory protein that affects the transformation of anti-inflammatory macrophage M2 phenotype. Upregulation of NPM1 expression is closely related to poor cardiac prognosis in MI patients. Specific knockout of macrophage NPM1 can promote the transformation of M2 macrophage phenotype and improve cardiac function and tissue repair after MI. Marginal zone B and B1 cell-specific protein 1 (Mzb1) is an endoplasmic reticulum resident protein that plays an important role in immune response by enhancing the interaction of Ig weight chains with chaperone glucose-regulated protein (GRP)94 and increasing IgM secretion (95). We have recently shown that Mzb1 expression is significantly reduced in AMI mice, while Mzb1 overexpression can attenuate ROS production, inflammation, mitochondrial dysfunction and myocardial apoptotic cells, and improve cardiac function associated with reduced inflammatory macrophage infiltration (96). Considering the important role of Mzb1 in the regulation of inflammation and immune cells, we speculate that Mzb1 may promote cardiac repair after MI by facilitating reparative macrophage phenotype.

4.2 Ventricular remodeling with diabetes cardiomyopathy (DCM)

Diabetes mellitus (DM) is a metabolic disease characterized with increased blood level of glucose due to the impairment of insulin production and action. Long-term diabetes with uncontrolled high blood glucose often causes serious microvascular complications in vital organs, such as the heart, pulmonary and kidney (97–99). DCM is one of the most serious complications in the patients with diabetes. DCM patients have significant structural and functional abnormalities in the heart muscle as well as heart ventricular remodeling (100). These abnormalities include left ventricular hypertrophy and fibrosis, extracellular matrix (ECM) deposition, intracellular lipid accumulation with cardiac systolic and diastolic dysfunction (101).

The mechanisms by which DM promotes the development of DCM are complex and not fully understood. Multiple factors including hyperglycemia, hyperlipidemia, insulin resistance, ROS and inflammation contribute to diabetic heart injury and dysfunction (102). Among them, inflammation is absolutely necessary for the development and progression of DCM. Diabetes

is a chronic low grade inflammatory disease, and metabolic disturbances, such as hyperglycemia and hyperlipidemia, stimulate immune cells infiltration into fat, vascular wall, the heart and kidney. These immune cells, especially macrophages, release inflammatory cytokines to initiate a low-grade inflammation in the infiltrated tissues or organs (103). Geng et al. (104) have reported that high glucose can directly induce macrophage activation and release M1 inflammatory cytokines through stimulating mitochondria ROS-dependent STING pathway *in vitro*. In the early stage of DCM, metabolic disturbance induces immune cell infiltration and chronic inflammatory response in the heart but does not significantly impair heart structure and function. However, continuous metabolic and inflammatory stress, especially inflammatory cytokines such as TNF α , IL-1 β released from activated macrophages, could induce oxidative stress, endoplasmic reticulum stress, mitochondria dysfunction in the heart, leading to cardiomyocyte injury, apoptosis, cardiac fibrosis, eventually develop the advanced DCM (105).

Macrophages have been shown to play a critical role in the development and progression of DCM (105). In the early stage of diabetes, macrophages are recruited into adipose tissues. These recruited macrophages release inflammatory cytokines including TNF α , IL-1 β and IL-6 to induce a local and systemic inflammation. The inflammatory cytokines inhibit insulin signaling pathway further to worsen systemic insulin resistance and metabolic disturbances (106). In addition, the inflammatory cytokines from systemic inflammation and released by infiltrated macrophages in the heart may initiate and exacerbate cardiac injury (107).

In addition to inflammatory cytokines, the impairment of macrophage phagocytosis may be another mechanism for DM development of DCM complication. Normally, macrophages eat up apoptotic cells and cellular debris by efferocytosis. It has been found that the phagocytic activity of macrophages in diabetic rat are impaired, which manifest low level of lysosomal enzymes and chemotactic activity in macrophages (108, 109). The impaired phagocytosis of macrophage is highly correlated with elevated blood glucose level, and the agents that lower blood glucose can improve macrophage phagocytosis (110). Thus, metabolic stress in DM may induce M1 macrophage activation to initiate cardiac inflammation to induce heart injury. On the other hand, hyperglycemia can also impair phagocytosis of M2 macrophages to impede clearance of apoptotic cells and cellular debris and the resolution of inflammation. Thus, the imbalance between M1 and M2 macrophages in microenvironment of heart tissue may importantly contribute to DCM (1, 106, 110, 111). The restoration of this balance may be an important therapeutic strategy for DCM. It has been shown that in streptozotocin (STZ) -induced diabetes models, the genetic deletion or pharmacological inhibition of chemokine receptor CCR2 suppresses M1 macrophage and slows down cardiac fibrosis. Additionally, metformin, the most popular medicine for clinical treatment of DM, has been shown to have selective inhibition of monocytes differentiation into M1 macrophages, and promotion of M2 macrophage to reduce inflammation and beneficial effects on the heart (112).

4.3 Heart failure

Heart failure (HF) refers to a clinical syndrome caused by various etiologies that impairs the structure and function of the heart, resulting in the heart being unable to pump enough blood to meet tissue metabolic needs (113). Based on ejection fraction, HF can be classified into two categories: HF with reduced ejection fraction (HFrEF) and HF with preserved ejection fraction (HFpEF). HFrEF exhibits cardiac systolic dysfunction, while HFpEF mainly presents cardiac diastolic dysfunction without significant reduction in ejection fraction (114). Although etiologies of HF are complex and heterogeneous, HF patients exhibit some common pathophysiological characteristics, such as the elevation of ventricular filling pressure and wall tension, and decreased myocardial contractile and ventricular compliance (115). Reduction in cardiac output may trigger systemic and local neurohumoral reflexes, immune response and other factors that cause the proliferation or apoptosis of myocardial cells, ECM deposition, inflammation, immune cell infiltration and myocardial fibrosis, resulting in ventricular pathological remodeling (11, 116, 117).

Regardless of the primary diseases of HF, inflammation and immune cell infiltration, especially monocytes/macrophage infiltration, play a pivotal role in the pathogenesis of HF (103). For HF caused by myocarditis or myocardial infarction, inflammation is the primary and essential factor to promote heart injury. Other types of HF, for example, hypertensive HF induced by aortic constriction, have significant increase in the infiltration of CCR2⁺ macrophages in myocardial tissue, as well as increased levels of local or circulating inflammatory cytokines, such as TNF α , IL-1 β , IL-6 and MCP-1, suggesting that HF increases circulating monocyte-derived CCR2⁺ inflammatory macrophages in the heart (55, 118, 119). Cardiac macrophages have both beneficial and detrimental effects in the development of HF. On the one hand, CRMs, specifically CCR2⁻ CRMs can clear up the dead cells or cellular debris to maintain heart homeostasis through endocytosis (77, 78). On the other hand, CRMs release/produce inflammatory cytokines or ROS which induce myocardial cell injury, apoptosis and cardiac injury and fibrosis, resulting in pathological ventricular remodeling and further worsening heart dysfunction (120, 121). Heart fibrosis may also alter and reshape heart electrophysiologic activity, and become the pathological basis for the formation of arrhythmias. However, HF often shifts CRMs into inflammatory M1 macrophages and produces large amounts of inflammatory cytokines and chemokines associated with impairment of macrophage endocytosis (122). Clinical and experimental studies have shown that there is a significant correlation between plasma inflammatory cytokine level and the severity of HF. Inflammatory cytokines, such as TNF α and IL-6, have been used for biomarkers to predict the progression of HF (122, 123). Specific depletion of macrophage reduces cardiac inflammation, attenuates cardiac hypertrophy, fibrosis and ventricular function in animal models of HF.

It should be pointed out that the most current studies on inflammation, M1/M2 macrophages and HF are based on HFrEF

(124). HFpEF exhibits more pronounced systemic inflammation and activated fibrotic pathway in the heart with increased profibrotic M2 macrophages. However, unlike anti-inflammatory beneficial effects of M2 macrophages in HFrEF, M2 macrophages in HFpEF exhibit more detrimental profibrotic effects. These M2 macrophages secrete large amounts of profibrotic factors such as TGF- β to enhance ventricular fibrosis, reduce ventricular compliance and impair ventricular relaxation (125, 126). Specific target macrophage phenotype is becoming a promising approach for treatment of HF, but it also faces many unsolved issues and enormous challenges.

4.4 Hypertensive left ventricular hypertrophy

Hypertension and its associated left ventricular hypertrophy (LVH) and HF remain a major challenge in the field of public health (127). Long-term high blood pressure causes disturb hemodynamic stress on the vascular and left ventricular wall, which promotes LVH hypertrophy, ECM deposition and fibrosis. Short-term ventricular hypertrophy and increased ECM is adaptive and may help to normalize ventricular wall stress and oxygen demand, which have compensatory effects on cardiac function (128). However, long-term ventricular hypertrophy, which lead to myocardium inflammation, fibrosis, cell death and development of HF, are unfavorable (129).

Increasing evidence has shown that the immune cells have been linked to the development and progression of hypertension and LVH (130). Long-term hemodynamic stress induces vascular and heart ventricular inflammatory responses that are increasingly recognized as key modulators of heart ventricular remodeling (131, 132). It has been reported that there is large number of monocytes and macrophages recruitment associated with increased expression of M1 macrophage cytokines in the heart and vascular wall of various hypertensive animal models (133, 134). Recruited macrophages release ROS and inflammatory cytokines, such as TNF α and IL-1 β , inducing vascular and heart inflammation, cell proliferation and ECM deposition, and thus promoting heart ventricular remodeling (11). TNF α is an inflammatory cytokine mainly produced by macrophage. We have shown that deficiency in the TNF α gene can lower blood pressure, improve endothelial function, and attenuate left ventricular hypertrophy and fibrosis accompanied by reduced macrophage infiltration in the heart and vessel of deoxycorticosterone acetate (DOCA)/salt hypertensive mice (134). Kain et al. (135) have reported that depletion of monocytes/macrophages using clodronate liposomes reduces LVH by 17% and cardiac fibrosis by 75% in aortic constriction and pressure overload hypertensive mice. We used clodronate liposomes to deplete macrophages and found that clodronate liposomes reduce circulating monocytes by 70%, and display remarkable reduction in vascular and ventricular macrophages, lowering blood pressure, vascular and heart hypertrophy and fibrosis in both angiotensin (Ang) II hypertensive mice and Dahl salt-sensitive hypertensive rats (136, 137). These results suggest that

macrophages in the heart and vessels play a critical role in hypertension and hypertensive heart remodeling (135).

Hypertension is often associated with increased circulating M1 inflammatory macrophages and expression of tissue M1 macrophage markers (138, 139). The mechanism of M1 macrophage activation in hypertension is not fully understood. The activation of the renin angiotensin system (RAS) has implicated in hypertension and hypertensive target end organ damage (140). Ang II is a pro-inflammatory mediator, Ang II promotes monocyte/macrophage infiltration into the vascular and heart wall. It has been shown that macrophages express all components of RAS (141). Recently we have shown that Ang II increases the expression of M1 pro-inflammatory cytokines TNF α , IL-1 β , and iNOS in human THP1/macrophage, the underlying mechanisms are mediated by the activation of HIF1 α /TLR4/NF- κ B signaling pathway, because siRNA HIF1 α prevents Ang II-induced macrophage activation (133). Furthermore, we show that expression of angiotensin type 1 receptor (AT1R) is increased in the peritoneal macrophage of DOCA/salt hypertensive mice, and specific knockout of AT1R in myeloid cells attenuates macrophage recruitment in vascular wall and reduces heart and vascular hypertrophy and fibrosis without significant reduction in blood pressure (133). These results suggest that RAS may regulate macrophage phenotype and polarization, and dysregulation of RAS in immune cells may be an important mechanism of inducing macrophage activation and the vascular and heart remodeling in hypertension, which warrants further investigation.

4.5 COVID-19 infection-induced myocardial injury

COVID-19 originally surfaces as an acute viral respiratory syndrome caused by acute respiratory syndrome coronavirus 2 (SARS-CoV2) infection (142). However, severe COVID-19 is often associated with acute cardiac injury and heart ventricular remodeling, manifested arrhythmias, myocardial ischemia, ventricular dysfunction, recurrent decompensated HF, or myocarditis (143). The mechanism of heart injury caused by COVID-19 is complex and not yet fully understood. The cardiac complications of COVID-19 are heterogeneous and can present at any periods of the COVID-19 infection, even occur as a sequela of post-infection (144). The patients with a history of heart disease and risk factors are more susceptible to have cardiac complications of COVID-19, or develop into more severe illness, even die (145). Serum level of troponin, as a biomarker of acute heart injury, increases in the patients with severe COVID-19, and the serum concentration of troponin is significantly correlated with the severity of the disease and the risk of death in the patients with COVID-19 (146).

Cardiac autopsy from the patients with COVID-19 has shown that, unlike traditional viral myocarditis which mainly involves lymphocytic infiltration, there are more CD68⁺ cells in the heart of the patients with COVID-19, and CD68⁺ cells are widely distributed in the myocardium and epicardium of the heart, suggesting an important role of macrophages in COVID-19 infection-induced cardiac injury (147). In addition, high levels of TNF α , IL-6, and

CCL2 inflammatory cytokines are detected in the circulating blood and the heart of patients with COVID-19, it is worth noting that these inflammatory cytokines are also important markers of M1 macrophage activation (148). These suggest that monocyte/macrophage infiltration might be more closely related to heart injury caused by SARS CoV-2 infection. Thus, some investigators propose that macrophages may play a more important role in myocarditis or cardiac injury caused by SARS-CoV-2 (149). Severe COVID-19 patients have long-term adverse heart remodeling, a group of autopsy data from the University of Alabama showed that 80% of the dead patients with severe COVID-19 had myocardial fibrosis, 72% had myocardial hypertrophy, and 66% had microembolization (150).

It is known that SARS-CoV2 enters host cells mainly through the interaction between angiotensin converting enzyme (ACE)2 receptor on the host cells and spike protein from SARS-CoV2 (151). After spike protein binds to ACE2 receptor on host cells, the virus is internalized and enters the cell, replicating DNAs within the host cells, which may disrupt the organelles, such as mitochondria and endoplasmic reticulum (152, 153). ACE2 is a component of RAS and plays an important role in the regulation of cardiovascular function. ACE2 receptor is widely distributed in the cardiovascular system (154). Thus, the ACE2 receptor may be an important target for SARS-CoV2 affecting the cardiovascular system. Unlike cardiac cells with rich ACE2 receptor, monocytes and macrophages have no or low level of ACE2 receptor, it is found that SARS-CoV2 enters and activates monocyte/macrophage mainly through GRP78 rather than ACE2 receptor (155). It is similar to the finding that is seen in macrophage activation syndrome: The activated macrophages in COVID-19 release large amount of inflammatory cytokines in their infiltrated organs, such as the lung and the heart, to trigger inflammatory storm, which may lead to fatal damage to the heart, lung and other vital organs (148, 156). Specific targets on macrophage phenotypes and reduction in the risk of the inflammatory storm may be a promising therapy for severe COVID-19 patients.

4.6 Heart valve diseases

Heart valve diseases are a common cardiovascular disease, which can be classified into congenital and acquired heart valve diseases (157). Due to significant alterations in cardiac hemodynamics within heart chambers, heart valve diseases often cause ventricular hypertrophy, fibrosis, and other ventricular remodeling as well as cardiac dysfunction (158). Myxomatous valve disease and calcific valve disease are the two most common types of heart valve disease. Myxomatous valve disease often invades the mitral valve, and the pathological changes mainly exhibit progressive extracellular matrix abnormalities and thickening of leaflets, which can ultimately lead to mitral prolapse and regurgitation (159). Aortic valve calcification is characterized by aortic stenosis and mineralization, which are more common in elderly and ischemic heart diseases (160).

Like myocardial tissue, there are multiple immune cells present in the development of heart valves, which account for

approximately 8% of the total number of heart cells. Among them, 75% of cells express the biomarkers of macrophages (161). Macrophage lineages in the heart valve are heterogeneous, including embryonic resident macrophages and macrophages derived from circulating monocytes. These macrophages play a critical role in heart valve development, physiological homeostasis, and pathological heart valve remodeling (162). During embryonic development, the heart valve is mainly composed of resident macrophages that express CCR2. After birth, macrophages from the hematopoietic lineage start to recruit to the heart valve (161, 163). In neonatal heart valves, macrophages mainly express M2 macrophage marker CD206, while in myxomatous heart valves, recruitment macrophage expressing M1 inflammatory markers significantly increases. Interestingly, the time and number of macrophage recruitment to the heart valve are closely related to the pathological thickening of the myxomatous heart valve (164). It has been shown that the heart valves have already recruited with large numbers of inflammatory macrophages before the heart valve begins to become pathological thickness and heart functional damage. The depletion of macrophages prevents heart valve thickness (163). These results suggest that macrophages play an important role in pathological remodeling of heart valves, especially myxomatous mitral valves.

4.7 Atrial fibrillation and heart electric remodeling

Atrial fibrillation (AF) is the most common type of cardiac arrhythmia associated with a range of heart diseases, such as ischemic coronary heart disease and HF (165). AF is often accompanied with electric remodeling and structure remodeling in atrial myocardial cells (166). Electrical remodeling mainly manifests as changes in the number, distribution and functional activity of atrial ion channels and gap junction proteins, resulting in a shortened or prolonged effective refractory period (167). The characteristics of structural remodeling are progressive collagen deposition and fibrosis in the atrium (168). Electric remodeling is considered an important factor affecting individual susceptibility to AF, whereas structural remodeling is a pathological base for persistent AF and other arrhythmias (169).

The increasing evidence suggests a close correlation between AF and inflammation. It is found that the number of immune cells expressing CD45⁺ and CD68⁺ (a macrophage marker) significantly increases in the atrium of AF patients, except macrophages, other immune cells, including mast cell, dendritic cells and CD3⁺ T cells, are also increased in the patients with AF (170, 171). The patients with AF have higher plasma level of inflammatory cytokines TNF α , IL-1 β and IL-6 (172). The immunotherapies that target on specific cytokines or their receptors have shown to exhibit therapeutic beneficial effects on AF. Colchicine is a broad-spectrum of anti-inflammatory drug that inhibits IL-1 β production and TNF α -induced NF- κ B inflammatory pathway. COPPS trial shows that treatment with colchicine for one month lowered the incidence of postoperative AF compared to placebo (173). As we know that IL-1 β is an important cytokine released by M1 macrophage, a clinical

trial demonstrates that targeting on IL-1 β by canakinumab can lower the incidence of AF recurrence after electrical cardioversion in the patients with heart diseases (174). In addition, two preclinical studies have shown that depletion of macrophages lowers the incidence of AF. These results strongly support the notion that inflammation and macrophages play an important role in the pathogenesis of AF (168). It should be noted that due to limited size of these clinical trials, the effectiveness of these anti-inflammatory drugs in treating AF still needs further validation through large-scale clinical trials.

Recent studies have shown that inflammation and immune system have great impact on atrial electric activity. As AF is always associated with cardiac chronic inflammation and immune cell infiltration, especially macrophages (168). It has been shown that inflammatory cytokines released by macrophages, such as TNF α , IL-1 β , IL-6 and macrophage inhibitor factor (MIF), can affect the activity of various ion channels in the atrium and induce atrial electric remodeling (175). TNF α inhibits T-type calcium channel α 1G subunit and sarcoplasmic reticulum Ca-ATPase that disrupt intracellular Ca₂⁺ homeostasis in atrial myocytes, enhancing arrhythmogenic activity (176). Macrophage-derived IL-1 β suppresses the expression of L-type calcium channel and decreases calcium channel expression, resulting in abnormal Ca²⁺ handling imbalance (76). Connexin proteins are important components of gap junction in myocardial cells and play a crucial role in the electric conduction of the heart, it has been shown that pro-inflammatory cytokines TNF α , IL-1 β and MIF can inhibit the expression of Cx40 and Cx43, affecting electric activity and conduction in the atrium (177). Therefore, immune cells or macrophages induce atrial electric remodeling through direct or indirect manners, promoting the occurrence of AF.

5 Macrophage-based therapeutic strategies for heart diseases

Despite significant progress in treatment options, cardiovascular diseases, such as MI and chronic HF, remain the leading cause of death over the world. Chronic inflammation and its related irreversible fibrosis are the main underlying pathophysiological causes that promote heart diseases and ventricular remodeling (178, 179). Cardiac macrophages have been recognized as the main driving force behind the development of these pathophysiological conditions. Macrophage-based immunotherapies have been recommended and may bring new prospects for the treatment of ventricular remodeling in heart diseases (119).

5.1 Inhibition of macrophage recruitment and the depletion of cardiac macrophages

As discussed above, recruited monocyte-derived macrophages importantly contribute to heart inflammation and adverse heart remodeling in many cases of heart diseases (54). Thus, the strategies aiming at inhibition cardiac macrophage recruitment or the

depletion of macrophages may be promising therapy for heart diseases. MCP-1, also known as CCL2, and its receptor CCR2 (CCL2-CCR2) axis is an important factor to induce the recruitment of monocytes and macrophages into the vascular wall and the heart (180). Recent several preclinical studies have shown that the inhibition of CCL2 through anti-CCL2 antibody or CCR2 depletion can attenuate adverse left ventricular remodeling in MI mice, and silencing CCR2 gene through siRNA reduces inflammatory monocytes/macrophages recruitment in ischemic areas and improves heart function in MI mice (181). Depletion of monocytes/macrophages have shown to have beneficial effects on the heart by the inhibition of deteriorating inflammatory responses. Kain et al. (135) have shown that the depletion of macrophage by clodronate liposomes significantly attenuates left ventricular hypertrophy and fibrosis and inhibits adverse cardiac remodeling in hypertensive animal models.

5.2 Therapeutic approaches for targeting macrophage phenotypes

Cardiac macrophages exhibit significant heterogeneity and plasticity. In general, monocyte-derived cardiac macrophage (CCR2⁺ CRM) are primarily linked to inflammatory response and pathogenic conditions, while yolk embryonic-derived CRMs (CCR2⁻ CRM) have cardioprotective effects and participate in heart reparative process (49, 182). The depletion of the entire cardiac macrophage population may result in compromised heart remodeling, due to the loss of reparative macrophages. Therefore, maintaining a delicate balance between protective and inflammatory macrophages through selective targets on macrophage phenotype is a highly attractive therapeutic strategy for treatment of heart diseases (12, 183). With the development of new biotechnologies, targeted therapies for macrophage phenotype are becoming possible.

Macrophage phenotype can be induced by cytokines, IL-1 β is not only a biomarker of inflammatory macrophage but also an important cytokine to induce inflammatory macrophage differentiation (1). It has been shown that treatment with anti-IL-1 β antibody or IL-1 β receptor antagonist canakinumab can significantly reduce inflammation and inhibit myocardial cell apoptosis in MI animal models, while treatment with IL-4 promotes the switching of inflammatory macrophage into reparative macrophage during the inflammatory phase of AMI, thus reducing infarct size and improving pump function of the heart (184, 185).

It should be pointed out that the selective targeting or reprogramming cardiac macrophage *in vivo* for treatment of heart diseases faces many challenges, as there is a lack of unique marker to distinguish cardiac macrophage phenotypes, making it difficult to accurately target subtypes of cardiac macrophages and avoid off target effects caused by attacking other immune cells. Using *ex vivo* reprogramming autologous macrophages may be another option to

overcome these shortcomings (186). For example, macrophages can be reprogrammed by incubating with M2 stimulators such as IL-10, IL-4 or TGF- β , and these reprogrammed macrophages have specific functions such as anti-inflammation, promoting angiogenesis and tissue repair, inhibiting cardiomyocyte apoptosis (187–189). Especially, autologous macrophage reprogramming *ex vivo* not only provide personalized therapy for patients, but also avoid off target macrophages. It has been demonstrated that transplantation of reprogrammed reparative macrophages induced by IL-4 or IL-10 can provide superior therapeutic benefits to the heart in mouse models of doxorubicin-induced HF and MI (190). In addition, induced pluripotent stem cell (iPSC)-derived macrophages can be generated from healthy donors, reprogramming iPSC-derived macrophages toward an anti-inflammatory phenotype are considered more effective option for treating heart diseases (191).

5.3 New therapeutic strategies for heart diseases based on macrophage drug delivery system

Recruitment of monocytes/macrophages and their associated inflammation are important pathogenesis of heart diseases. Since monocytes/macrophages have the ability to specifically migrate and recruit into the inflamed site, macrophages and macrophage-derived exosomes may serve as excellent drug delivery carriers (192). Nanoparticles (NPs) are formulations of diverse molecular compounds with complex architecture which are spatially organized on the scale of tens of nanometers (193). Unlike conventional pharmacotherapies with only a single therapeutic function, engineered multifunctional NPs can be designed to have ability to sense and deliver payloads, such as pharmacological activators, inhibitors, or siRNA (194). The combination of the various chemical properties of NPs and the biological targeting function of macrophages is becoming a new paradigm for the treatment of various diseases, including tumors, metabolic and cardiovascular diseases. Recently, Hu et al. (195) reported that they successfully developed a mannan-based Que@MOF/Man nanoparticles which are preferentially internalized by M2 macrophages through binding to their CD206 receptor. The administration of Que@MFO/Man exhibited favorable results in the resolution of inflammation, the prevention of adverse heart remodeling and the improvement of heart function in MI rats.

5.4 Clinical trials based on inhibition of macrophages or inflammation for treatment of heart disease

Current clinical trials of immunotherapy for heart diseases are mainly based on inflammatory cytokines, including IL-1 β , TNF α , IL-6 and CCL2 (196). These cytokines are important biomarkers for M1 macrophages, which are also the main source of these

inflammatory cytokines. Some anti-inflammatory medications or antibodies that focus on inhibiting or blocking these inflammatory cytokines have developed and clinically used for the treatment of heart diseases in clinical trials, such as MI and HF (197). A randomized and double-blind clinical trial has shown that long-term treatment with canakinumab, a therapeutic IL-1 β antibody, can successfully lower inflammation and rate of recurrent cardiovascular events in the patients with post-MI (198). Colchicine is a broad-spectrum of anti-inflammatory drug that inhibits IL-1 β production and TNF α -induced NF- κ B inflammatory pathway. Two clinical trials (COLCOT and LoDoCo2) have shown that colchicine significantly reduces ischemic cardiovascular events such as angina, MI and cardiac arrest in the patients with coronary artery diseases (199, 200). The ASSAIL-MI trial demonstrated that treatment with IL-6 antibody tocilizumab has tendency to reduce infarct size in MI patients (201). In addition, some ongoing clinical trials based on anti-inflammatory therapies have shown to reduce the incidence of major adverse cardiovascular events in the patients with coronary arterial diseases (NCT01906749 & NCT05130892) and MI (NCT02648464 & NCT01372930) (202, 203).

6 Conclusion and perspective

Macrophages, as a key type of immune cells residing in the heart, participate in regulating heart function in physiological condition, and induce heart remodeling in pathophysiological condition by altering their phenotypes and secreting a variety of cytokines or mediators (204). In a healthy heart, CRMs are predominated by the embryonic derived CCR2⁻ macrophages, which participate in the maintenance of heart homeostasis by clearing up apoptotic cells and cellular debris through endocytosis (77, 78). When the heart faces challenges, such as inflammation, ischemia and mechanical stress, which stimulate monocytes-derived CCR2⁺ CRMs recruitment in the heart, the cardiac macrophages undergo metabolic reprogramming to become pro-inflammatory M1 macrophage, and participate in heart ventricular remodeling under the pathological condition by releasing inflammatory cytokines and chemokines (3, 12, 111, 205). When the signal of inflammation or environment stress weakens, inflammatory M1 macrophages transform into anti-inflammatory M2 macrophages to promote heart repair.

Despite significant advancement, there are still some limitations and challenges in the field of cardiac macrophages. Firstly, there is currently no specific biomarker that can distinguish beneficial and harmful macrophage phenotypes. Current biomarkers are mainly based on cytokines and chemokines secreted by macrophages, such as CCL2, CCR2, TNF α , IL-1, IL-4 and IL-10 (119). CCR2 is the mostly used as a biomarker for distinguishing inflammatory and resident cardiac macrophages, but it is not a good molecular marker for distinguishing beneficial or detrimental macrophages (161). Some anti-inflammatory drugs or antibodies developed based on these biomarkers have entered clinical trials for immunotherapy of heart disease patients (206). However, these drugs or biological reagents cannot precisely target harmful macrophage phenotype,

therefore these therapies have uncertain efficacy and side effects. Next, although the strategy targeting pathological-related cardiac macrophages or regulating function of cardiac macrophage have enormous therapeutic potential, there is currently a lack of an effective drug delivery system that specifically targets tissue or organ macrophages, especially cardiac macrophages. Finally, the current models used for *in vitro* and *in vivo* experiments of cardiac macrophages are too simple to simulate the complex pathological conditions of cardiac macrophage heterogeneity and plasticity in the patients, making it difficult to translate experimental results into clinical practice (207).

Overcoming these difficulties and challenges is our main tasks for future research on cardiac macrophages. 1) With advancement in single cell RNA-seq (scRNA-seq) and spatiotemporal transcriptomics, scRNA-seq can accurately detect and classify cell subsets, discover new cell population, and provide transcriptome profiles of cell subsets rather than just a few biomarkers (208). Spatiotemporal transcriptomics can analyze spatially separated genetic information from individual cells, and preserve their spatial location and interactions under normal heart and various pathological conditions of heart diseases (209). The application of these advanced biological technologies will enable us to precisely determine the subsets of cardiac macrophages and identify specific molecular targets on cardiac macrophages for heart diseases, particularly identifying specific molecular markers that distinguish beneficial and harmful cardiac macrophages. 2) The combination of nanoparticle drug delivery with *ex vivo* macrophage reprogramming can overcome the drawback of macrophage drug delivery systems. Nanoparticles carrying drugs, bioactive substances, or siRNA can also achieve precise targeting of cardiac macrophages, regulate macrophage function *in vivo* to achieve the goal of treating heart disease (195). Therefore, developing advanced drug delivery strategies that precisely target macrophages is expected to bring the vision of successful clinical translation to reality. 3) Long term *in vivo* studies using large animal models or humanized mice can address the translation gap between small rodents, and humans, and explore the role and phenotypic changes of macrophages over time in different cardiac pathological setting. In addition, using a culture model of precision heart slices (living myocardial slices) *ex vivo* to investigate cardiac macrophage can overcome the problem of current *in vitro* macrophage culture model being too simple (210).

It is worth noting that macrophage targeted therapies are still in the early stages of development, and these therapeutic strategies require a comprehensive evaluation of their efficacy and safety in heart disease patients. Further large-scale clinical trials may need to translate these fundamental findings into clinical practice. However, these promising therapeutic strategies are paving the way for a new era of personalized medicine based on cellular therapy, which may be applied to treat other human inflammatory diseases.

Author contributions

MK: Writing – original draft, Conceptualization. HJ: Writing – original draft, Conceptualization. MF: Writing – original draft. HR:

Writing – original draft, JG: Writing – original draft, YL: Writing – original draft, LZ: Writing – original draft, Funding acquisition, Conceptualization, M-SZ: Writing – review & editing, Writing – original draft, Funding acquisition, Conceptualization.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the grants from the National Natural Science Foundation of China (Nos. 82270434, 81970357) to M-SZ, and the National Natural Science Foundation of China (82100316), the Doctoral Scientific Research Foundation of Liaoning Province (2022-BS-342), Scientific research fund Project of Education Department of Liaoning Province (LJKZ1135) to LZ.

References

- Shapouri-Moghaddam A, Mohammadian S, Vazini H, Taghadosi M, Esmaili SA, Mardani F, et al. Macrophage plasticity, polarization, and function in health and disease. *J Cell Physiol.* (2018) 233:6425–40. doi: 10.1002/jcp.26429
- Sheu KM, Hoffmann A. Functional hallmarks of healthy macrophage responses: their regulatory basis and disease relevance. *Annu Rev Immunol.* (2022) 40:295–321. doi: 10.1146/annurev-immunol-101320-031555
- Rodríguez-Morales P, Franklin RA. Macrophage phenotypes and functions: resolving inflammation and restoring homeostasis. *Trends Immunol.* (2023) 44:986–98. doi: 10.1016/j.it.2023.10.004
- Locati M, Curtale G, Mantovani A. Diversity, mechanisms, and significance of macrophage plasticity. *Annu Rev Pathol.* (2020) 15:123–47. doi: 10.1146/annurev-pathmechdis-012418-012718
- Yunna C, Mengru H, Lei W, Weidong C. Macrophage M1/M2 polarization. *Eur J Pharmacol.* (2020) 877:73090–8. doi: 10.1016/j.ejphar.2020.173090
- Kadomoto S, Izumi K, Mizokami A. Macrophage polarity and disease control. *Int J Mol Sci.* (2021) 23:144–56. doi: 10.3390/ijms23010144
- Epelman S, Lavine KJ, Beaudin AE, Sojka DK, Carrero JA, Calderon B, et al. Embryonic and adult-derived resident cardiac macrophages are maintained through distinct mechanisms at steady state and during inflammation. *Immunity.* (2014) 40:91–104. doi: 10.1016/j.immuni.2013.11.019
- Zhao Y, Zou W, Du J, Zhao Y. The origins and homeostasis of monocytes and tissue-resident macrophages in physiological situation. *J Cell Physiol.* (2018) 233:6425–39. doi: 10.1002/jcp.26461
- Qin D, Zhang Y, Liu F, Xu X, Jiang H, Su Z, et al. Spatiotemporal development and the regulatory mechanisms of cardiac resident macrophages: Contribution in cardiac development and steady state. *Acta Physiol (Oxf).* (2024) 240:e14088–103. doi: 10.1111/apha.14088
- Mouton AJ, Li X, Hall ME, Hall JE. Obesity, hypertension, and cardiac dysfunction: novel roles of immunometabolism in macrophage activation and inflammation. *Circ Res.* (2020) 126:789–806. doi: 10.1161/circresaha.119.312321
- Wong NR, Mohan J, Kopecky BJ, Guo S, Du L, Leid J, et al. Resident cardiac macrophages mediate adaptive myocardial remodeling. *Immunity.* (2021) 54:2072–88. doi: 10.1016/j.immuni.2021.07.003
- Sun X, Li Y, Deng Q, Hu Y, Dong J, Wang W, et al. Macrophage polarization, metabolic reprogramming, and inflammatory effects in ischemic heart disease. *Front Immunol.* (2022) 13:934040–6. doi: 10.3389/fimmu.2022.934040
- Orecchioni M, Ghosheh Y, Pramod AB, Ley K. Macrophage Polarization: Different Gene Signatures in M1(LPS+) vs. Classically and M2(LPS-) vs. Alternatively Activated Macrophages. *Front Immunol.* (2019) 10:1084–97. doi: 10.3389/fimmu.2019.01084
- Huang X, Li Y, Fu M, Xin HB. Polarizing macrophages *in vitro*. *Methods Mol Biol.* (2018) 1784:119–26. doi: 10.1007/978-1-4939-7837-3_12
- Lauterbach MA, Hanke JE, Serefidou M, Mangan MSJ, Kolbe CC, Hess T, et al. Toll-like receptor signaling rewires macrophage metabolism and promotes histone acetylation via ATP-citrate lyase. *Immunity.* (2019) 51:997–1011. doi: 10.1016/j.immuni.2019.11.009
- Zhu L, Zhao Q, Yang T, Ding W, Zhao Y. Cellular metabolism and macrophage functional polarization. *Int Rev Immunol.* (2015) 34:82–100. doi: 10.3109/08830185.2014.969421
- Ruytinx P, Proost P, Van Damme J, Struyf S. Chemokine-induced macrophage polarization in inflammatory conditions. *Front Immunol.* (2018) 9:1930–41. doi: 10.3389/fimmu.2018.01930
- Ye L, Jiang Y, Zhang M. Crosstalk between glucose metabolism, lactate production and immune response modulation. *Cytokine Growth Factor Rev.* (2022) 68:81–92. doi: 10.1016/j.cytogfr.2022.11.001
- Huang SC, Everts B, Ivanova Y, O'Sullivan D, Nascimento M, Smith AM, et al. Cell-intrinsic lysosomal lipolysis is essential for alternative activation of macrophages. *Nat Immunol.* (2014) 15:846–55. doi: 10.1038/ni.2956
- Funes SC, Rios M, Escobar-Vera J, Kalergis AM. Implications of macrophage polarization in autoimmunity. *Immunology.* (2018) 154:186–95. doi: 10.1111/imm.12910
- Han JM, Lee EK, Gong SY, Sohng JK, Kang YJ, Jung HJ. Sparassis crispa exerts anti-inflammatory activity via suppression of TLR-mediated NF- κ B and MAPK signaling pathways in LPS-induced RAW264.7 macrophage cells. *J Ethnopharmacol.* (2019) 231:10–8. doi: 10.1016/j.jep.2018.11.003
- Wei YY, Fan YM, Ga Y, Zhang YN, Han JC, Hao ZH. Shaoyao decoction attenuates DSS-induced ulcerative colitis, macrophage and NLRP3 inflammasome activation through the MKP1/NF- κ B pathway. *Phytomedicine.* (2021) 92:153743–58. doi: 10.1016/j.phymed.2021.153743
- Zhao D, Zhang X, Feng Y, Bian Y, Fu Z, Wu Y, et al. Taurine alleviates LPS-induced acute lung injury by suppressing TLR-4/NF- κ B pathway. *Adv Exp Med Biol.* (2022) 1370:63–72. doi: 10.1007/978-3-030-93337-1_6
- Matveichuk OV, Ciesielska A, Hromada-Judycha A, Nowak N, Ben Amor I, Traczyk G, et al. Flotillins affect LPS-induced TLR4 signaling by modulating the trafficking and abundance of CD14. *Cell Mol Life Sci.* (2024) 81:191–215. doi: 10.1007/s00018-024-05221-3
- Regdon Z, Robaszkiewicz A, Kovács K, Rygielska Ż, Hegedűs C, Bodoor K, et al. LPS protects macrophages from AIF-independent parthanatos by downregulation of PARP1 expression, induction of SOD2 expression, and a metabolic shift to aerobic glycolysis. *Free Radic Biol Med.* (2019) 131:184–96. doi: 10.1016/j.freeradbiomed.2018.11.034
- Zhao H, Wu L, Yan G, Chen Y, Zhou M, Wu Y, et al. Inflammation and tumor progression: signaling pathways and targeted intervention. *Signal Transduct Target Ther.* (2021) 6:263–308. doi: 10.1038/s41392-021-00658-5
- Dichtl S, Lindenthal L, Zeitler I, Behnke K, Schlösser D, Strobl B, et al. Lactate and IL6 define separable paths of inflammatory macrophage adaptation. *Sci Adv.* (2021) 7:eabg3505–15. doi: 10.1126/sciadv.abg3505
- Liu Y, Xu R, Gu H, Zhang E, Qu J, Cao W, et al. Metabolic reprogramming in macrophage responses. *biomark Res.* (2021) 9:1. doi: 10.1186/s40364-020-00251-y
- Viola A, Munari F, Sánchez-Rodríguez R, Sclaro T, Castegna A. The metabolic signature of macrophage responses. *Front Immunol.* (2019) 10:1462–77. doi: 10.3389/fimmu.2019.01462
- Pradhan P, Vijayan V, Cirkseña K, Buettner FFR, Igarashi K, Motterlini R, et al. Genetic BACH1 deficiency alters mitochondrial function and increases NLRP3 inflammasome activation in mouse macrophages. *Redox Biol.* (2022) 51:102265–75. doi: 10.1016/j.redox.2022.102265
- Pecchillo Cimmino T, Ammendola R, Cattaneo F, Esposito G. NOX dependent ROS generation and cell metabolism. *Int J Mol Sci.* (2023) 24:2086–101. doi: 10.3390/ijms24032086

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

32. Kierans SJ, Taylor CT. Regulation of glycolysis by the hypoxia-inducible factor (HIF): implications for cellular physiology. *J Physiol.* (2021) 599:23–37. doi: 10.1113/jp280572
33. Gayatri MB, Kancha RK, Patchva D, Velugonda N, Gundeti S, Reddy ABM. Metformin exerts antileukemic effects by modulating lactate metabolism and overcomes imatinib resistance in chronic myelogenous leukemia. *FEBS J.* (2023) 290:4480–95. doi: 10.1111/febs.16818
34. Wang T, Liu H, Lian G, Zhang SY, Wang X, Jiang C. HIF1 α -induced glycolysis metabolism is essential to the activation of inflammatory macrophages. *Mediators Inflamm.* (2017) 2017:9029327–36. doi: 10.1155/2017/9029327
35. Masoumi M, Mehrabzadeh M, Mahmoudzahi S, Mousavi MJ, Jamalzei S, Sahebkar A, et al. Role of glucose metabolism in aggressive phenotype of fibroblast-like synoviocytes: Latest evidence and therapeutic approaches in rheumatoid arthritis. *Int Immunopharmacol.* (2020) 89:107064–71. doi: 10.1016/j.intimp.2020.107064
36. Doddapattar P, Dev R, Ghatge M, Patel RB, Jain M, Dhanesha N, et al. Myeloid cell PKM2 deletion enhances efferocytosis and reduces atherosclerosis. *Circ Res.* (2022) 130:1289–305. doi: 10.1161/circresaha.121.320704
37. Poels K, Schnitzler JG, Waissi F, Levels JHM, Stroes ESG, Daemen M, et al. Inhibition of PFKFB3 hampers the progression of atherosclerosis and promotes plaque stability. *Front Cell Dev Biol.* (2020) 8:581641–50. doi: 10.3389/fcell.2020.581641
38. Corcoran SE, O'Neill LA. HIF1 α and metabolic reprogramming in inflammation. *J Clin Invest.* (2016) 126:3699–707. doi: 10.1172/jci84431
39. Lv JJ, Wang H, Zhang C, Zhang TJ, Wei HL, Liu ZK, et al. CD147 sparks atherosclerosis by driving M1 phenotype and impairing efferocytosis. *Circ Res.* (2024) 134:165–85. doi: 10.1161/circresaha.123.323223
40. Wang T, Lu H. Ganoderic acid A inhibits ox-LDL-induced THP-1-derived macrophage inflammation and lipid deposition via Notch1/PPAR γ /CD36 signaling. *Adv Clin Exp Med.* (2021) 30:1031–41. doi: 10.17219/acem/137914
41. Zhou MS, Chadipiralla K, Mendez AJ, Jaimes EA, Silverstein RL, Webster K, et al. Nicotine potentiates proatherogenic effects of oxLDL by stimulating and upregulating macrophage CD36 signaling. *Am J Physiol Heart Circ Physiol.* (2013) 305:H563–74. doi: 10.1152/ajpheart.00042.2013
42. Dang B, Gao Q, Zhang L, Zhang J, Cai H, Zhu Y, et al. The glycolysis/HIF-1 α axis defines the inflammatory role of IL-4 primed macrophages. *Cell Rep.* (2023) 42:112471–90. doi: 10.1016/j.celrep.2023.112471
43. Junttila IS. Tuning the cytokine responses: an update on interleukin (IL)-4 and IL-13 receptor complexes. *Front Immunol.* (2018) 9:888–93. doi: 10.3389/fimmu.2018.00888
44. Jia DJ, Wang QW, Hu YY, He JM, Ge QW, Qi YD, et al. Lactobacillus johnsonii alleviates colitis by TLR1/2-STAT3 mediated CD206(+) macrophages(IL-10) activation. *Gut Microbes.* (2022) 14:2145843–61. doi: 10.1080/19490976.2022.2145843
45. Röszer T. Understanding the mysterious M2 macrophage through activation markers and effector mechanisms. *Mediators Inflamm.* (2015) 2015:816460–75. doi: 10.1155/2015/816460
46. Gianopoulos I, Daskalopoulou SS. Macrophage profiling in atherosclerosis: understanding the unstable plaque. *Basic Res Cardiol.* (2024) 119:35–56. doi: 10.1007/s00395-023-01023-z
47. Xue S, Su Z, Liu D. Immunometabolism and immune response regulate macrophage function in atherosclerosis. *Ageing Res Rev.* (2023) 90:101993–2003. doi: 10.1016/j.arr.2023.101993
48. Kieler M, Hofmann M, Schabbauer G. More than just protein building blocks: how amino acids and related metabolic pathways fuel macrophage polarization. *FEBS J.* (2021) 288:3694–714. doi: 10.1111/febs.15715
49. Zaman R, Epelman S. Resident cardiac macrophages: Heterogeneity and function in health and disease. *Immunology.* (2022) 55:1549–63. doi: 10.1016/j.immuni.2022.08.009
50. Yap J, Cabrera-Fuentes HA, Irei J, Hausenloy DJ, Boisvert WA. Role of macrophages in cardioprotection. *Int J Mol Sci.* (2019) 20:2474–91. doi: 10.3390/ijms20102474
51. Lavine KJ, Epelman S, Uchida K, Weber KJ, Nichols CG, Schilling JD, et al. Distinct macrophage lineages contribute to disparate patterns of cardiac recovery and remodeling in the neonatal and adult heart. *Proc Natl Acad Sci USA.* (2014) 111:16029–34. doi: 10.1073/pnas.1406508111
52. Chen R, Zhang S, Liu F, Xia L, Wang C, Sandoghchian Shotorbani S, et al. Renewal of embryonic and neonatal-derived cardiac-resident macrophages in response to environmental cues abrogated their potential to promote cardiomyocyte proliferation via Jagged-1-Notch1. *Acta Pharm Sin B.* (2023) 13:128–41. doi: 10.1016/j.apsb.2022.08.016
53. Dick SA, Wong A, Hamidzadeh H, Nejat S, Nechanitzky R, Vohra S, et al. Three tissue resident macrophage subsets coexist across organs with conserved origins and life cycles. *Sci Immunol.* (2022) 7:eabf7777–94. doi: 10.1126/sciimmunol.abf7777
54. Dick SA, Macklin JA, Nejat S, Momen A, Clemente-Casares X, Althagafi MG, et al. Self-renewing resident cardiac macrophages limit adverse remodeling following myocardial infarction. *Nat Immunol.* (2019) 20:29–39. doi: 10.1038/s41590-018-0272-2
55. Bajpai G, Bredemeyer A, Li W, Zaitsev K, Koenig AL, Lokshina I, et al. Tissue resident CCR2- and CCR2+ Cardiac macrophages differentially orchestrate monocyte recruitment and fate specification following myocardial injury. *Circ Res.* (2019) 124:263–78. doi: 10.1161/circresaha.118.314028
56. Varró A, Tomek J, Nagy N, Virág L, Passini E, Rodriguez B, et al. Cardiac transmembrane ion channels and action potentials: cellular physiology and arrhythmogenic behavior. *Physiol Rev.* (2021) 101:1083–176. doi: 10.1152/physrev.00024.2019
57. Nguyễn UC, Verzaal NJ, Van Nieuwenhoven FA, Vernooij K, Prinzen FW. Pathobiology of cardiac dyssynchrony and resynchronization therapy. *Europace.* (2018) 20:1898–909. doi: 10.1093/europace/euy035
58. Xie W, Gao S, Yang Y, Li H, Zhou J, Chen M, et al. CYLD deubiquitinates plakoglobin to promote Cx43 membrane targeting and gap junction assembly in the heart. *Cell Rep.* (2022) 41:111864–86. doi: 10.1016/j.celrep.2022.111864
59. Carmeliet E. Conduction in cardiac tissue. Historical reflections. *Physiol Rep.* (2019) 7:e13860–72. doi: 10.14814/phy2.13860
60. Ahir BK, Pratten MK. Structure and function of gap junction proteins: role of gap junction proteins in embryonic heart development. *Int J Dev Biol.* (2014) 58:649–62. doi: 10.1387/ijdb.140188dp
61. Johnson RD, Camelliti P. Role of non-myocyte gap junctions and connexin hemichannels in cardiovascular health and disease: novel therapeutic targets? *Int J Mol Sci.* (2018) 19:886–911. doi: 10.3390/ijms19030866
62. Dhein S, Salameh A. Remodeling of cardiac gap junctional cell-cell coupling. *Cells.* (2021) 10:2422–40. doi: 10.3390/cells10092422
63. Leybaert L, De Smet MA, Lissoni A, Allewaert R, Roderick HL, Bultynck G, et al. Connexin hemichannels as candidate targets for cardioprotective and anti-arrhythmic treatments. *J Clin Invest.* (2023) 133:e168117–29. doi: 10.1172/jci168117
64. Kreuzberg MM, Willecke K, Bukauskas FF. Connexin-mediated cardiac impulse propagation: connexin 30.2 slows atrioventricular conduction in mouse heart. *Trends Cardiovasc Med.* (2006) 16:266–72. doi: 10.1016/j.tcm.2006.05.002
65. Zhang M, Wang ZZ, Chen NH. Connexin 43 phosphorylation: implications in multiple diseases. *Molecules.* (2023) 28:4914–29. doi: 10.3390/molecules28134914
66. Xia R, Loy S, Käb S, Titova A, Schulz C, Massberg S, et al. Isolation and culture of resident cardiac macrophages from the murine sinoatrial and atrioventricular node. *J Vis Exp.* (2021) 171. doi: 10.3791/62236
67. Hulsmans M, Clauss S, Xiao L, Aguirre AD, King KR, Hanley A, et al. Macrophages facilitate electrical conduction in the heart. *Cell.* (2017) 169:510–22. doi: 10.1016/j.cell.2017.03.050
68. Simon-Chica A, Wülfers EM, Kohl P. Nonmyocytes as electrophysiological contributors to cardiac excitation and conduction. *Am J Physiol Heart Circ Physiol.* (2023) 325:H475–91. doi: 10.1152/ajpheart.00184.2023
69. Cox CB, Castro M, Brown TL, Bigley NJ. IL-10 and TGF- β Increase connexin-43 expression and membrane potential of HL-1 cardiomyocytes coupled with RAW 264.7 macrophages. *Immunohorizons.* (2022) 6:334–43. doi: 10.4049/immunohorizons.2100104
70. Nicolás-Ávila JA, Hidalgo A, Ballesteros I. Specialized functions of resident macrophages in brain and heart. *J Leukocyte Biol.* (2018) 104:743–56. doi: 10.1002/jlb.6mr0118-041r
71. Silvestre JS, Vandecasteele G. Heart rate: when macrophages hit the note. *Med Sci (Paris).* (2018) 34:820–3. doi: 10.1051/medsci/2018207
72. Calhoun PJ, Phan AV, Taylor JD, James CC, Padgett RL, Zeitz MJ, et al. Adenovirus targets transcriptional and posttranslational mechanisms to limit gap junction function. *FASEB J.* (2020) 34:9694–712. doi: 10.1096/fj.202000667R
73. Xu R, Li C, Wu Y, Shen L, Ma J, Qian J, et al. Role of KCa3.1 channels in macrophage polarization and its relevance in atherosclerotic plaque instability. *Arterioscler Thromb Vasc Biol.* (2017) 37:226–36. doi: 10.1161/atvbaha.116.308461
74. Billur D, Olgar Y, Durak A, Yozgat AH, Unay S, Tuncay E, et al. An increase in intercellular crosstalk and electrotonic coupling between cardiomyocytes and nonmyocytes reshapes the electrical conduction in the metabolic heart characterized by short QT intervals in ECGs. *Cell Biochem Funct.* (2023) 41:1526–42. doi: 10.1002/cbf.3893
75. Nasser MI, Zhu S, Huang H, Zhao M, Wang B, Ping H, et al. Macrophages: First guards in the prevention of cardiovascular diseases. *Life Sci.* (2020) 250:117559. doi: 10.1016/j.lfs.2020.117559
76. Sun Z, Zhou D, Xie X, Wang S, Wang Z, Zhao W, et al. Cross-talk between macrophages and atrial myocytes in atrial fibrillation. *Basic Res Cardiol.* (2016) 111:63–71. doi: 10.1007/s00395-016-0584-z
77. Razi S, Yaghmoorian Khojini J, Kargarijam F, Panahi S, Tahershamsi Z, Tajbakhsh A, et al. Macrophage efferocytosis in health and disease. *Cell Biochem Funct.* (2023) 41:152–65. doi: 10.1002/cbf.3780
78. Li Y, Li Q, Fan GC. Macrophage efferocytosis in cardiac pathophysiology and repair. *Shock.* (2021) 55:177–88. doi: 10.1097/shk.0000000000001625
79. Jin J, Wang Y, Liu Y, Chakrabarti S, Su Z. Cardiac resident macrophages: Spatiotemporal distribution, development, physiological functions, and their translational potential on cardiac diseases. *Acta Pharm Sin B.* (2024) 14:1483–93. doi: 10.1016/j.apsb.2023.12.018
80. Tucker NR, Chaffin M, Fleming SJ, Hall AW, Parsons VA, Bedi KC Jr., et al. Transcriptional and cellular diversity of the human heart. *Circulation.* (2020) 142:466–82. doi: 10.1161/circulationaha.119.045401

81. Kaier TE, Alaour B, Marber M. Cardiac troponin and defining myocardial infarction. *Cardiovasc Res.* (2021) 117:2203–15. doi: 10.1093/cvr/cvaa331
82. Bhatt AS, Ambrosy AP, Velazquez EJ. Adverse remodeling and reverse remodeling after myocardial infarction. *Curr Cardiol Rep.* (2017) 19:71–80. doi: 10.1007/s11886-017-0876-4
83. Lorusso R, Matteucci M, Lerakis S, Ronco D, Menicanti L, Sharma SK, et al. Postmyocardial infarction ventricular aneurysm: JACC focus seminar 5/5. *J Am Coll Cardiol.* (2024) 83:1917–35. doi: 10.1016/j.jacc.2024.02.044
84. Jia D, Chen S, Bai P, Luo C, Liu J, Sun A, et al. Cardiac resident macrophage-derived legumain improves cardiac repair by promoting clearance and degradation of apoptotic cardiomyocytes after myocardial infarction. *Circulation.* (2022) 145:1542–56. doi: 10.1161/circulationaha.121.057549
85. Peet C, Ivetic A, Bromage DI, Shah AM. Cardiac monocytes and macrophages after myocardial infarction. *Cardiovasc Res.* (2020) 116:1101–12. doi: 10.1093/cvr/cvz336
86. Yang L, Wang B, Zhou Q, Wang Y, Liu X, Liu Z, et al. MicroRNA-21 prevents excessive inflammation and cardiac dysfunction after myocardial infarction through targeting KBTBD7. *Cell Death Dis.* (2018) 9:769–82. doi: 10.1038/s41419-018-0805-5
87. Francisco J, Del Re DP. Inflammation in myocardial ischemia/reperfusion injury: underlying mechanisms and therapeutic potential. *Antioxidants (Basel).* (2023) 12:1944–74. doi: 10.3390/antiox12111944
88. Ma Y, Mouton AJ, Lindsey ML. Cardiac macrophage biology in the steady-state heart, the aging heart, and following myocardial infarction. *Transl Res.* (2018) 191:15–28. doi: 10.1016/j.trsl.2017.10.001
89. Kubota A, Frangogiannis NG. Macrophages in myocardial infarction. *Am J Physiol Cell Physiol.* (2022) 323:C1304–24. doi: 10.1152/ajpcell.00230.2022
90. Glington KE, Ma W, Lantz C, Grigoryeva LS, Deberge M, Liu X, et al. Macrophage-produced VEGFC is induced by efferocytosis to ameliorate cardiac injury and inflammation. *J Clin Invest.* (2022) 132:e140685–99. doi: 10.1172/jci140685
91. Cai S, Zhao M, Zhou B, Yoshii A, Bugg D, Villet O, et al. Mitochondrial dysfunction in macrophages promotes inflammation and suppresses repair after myocardial infarction. *J Clin Invest.* (2023) 133:e159498–511. doi: 10.1172/jci159498
92. Tan B, Shi X, Zhang J, Qin J, Zhang N, Ren H, et al. Inhibition of rspl-1g4 facilitates checkpoint blockade therapy by switching macrophage polarization. *Cancer Res.* (2018) 78:4929–42. doi: 10.1158/0008-5472.Can-18-0152
93. Huang CK, Dai D, Xie H, Zhu Z, Hu J, Su M, et al. Lgr4 governs a pro-inflammatory program in macrophages to antagonize post-infarction cardiac repair. *Circ Res.* (2020) 127:953–73. doi: 10.1161/circresaha.119.315807
94. Zhang S, Zhang Y, Duan X, Wang B, Zhan Z. Targeting NPM1 epigenetically promotes postinfarction cardiac repair by reprogramming reparative macrophage metabolism. *Circulation.* (2024) 149:1982–2001. doi: 10.1161/circulationaha.123.065506
95. Andreani V, Ramamoorthy S, Pandey A, Lupar E, Nutt SL, Lämmermann T, et al. Cochaperone Mzb1 is a key effector of Blimp1 in plasma cell differentiation and β 1-integrin function. *Proc Natl Acad Sci USA.* (2018) 115:E9630–9. doi: 10.1073/pnas.1809739115
96. Zhang L, Wang YN, Ju JM, Shabanova A, Li Y, Fang RN, et al. Mzb1 protects against myocardial infarction injury in mice via modulating mitochondrial function and alleviating inflammation. *Acta Pharmacol Sin.* (2021) 42:691–700. doi: 10.1038/s41401-020-0489-0
97. Zhang RH, Zhou JB, Cai YH, Shu LP, Simó R, Lecube A. Non-linear association between diabetes mellitus and pulmonary function: a population-based study. *Respir Res.* (2020) 21:292–300. doi: 10.1186/s12931-020-01538-2
98. Salvador DB Jr, Gamba MR, Gonzalez-Jaramillo N, Gonzalez-Jaramillo V, Raguindin PFN, Minder B, et al. Diabetes and myocardial fibrosis: A systematic review and meta-analysis. *JACC Cardiovasc Imaging.* (2022) 15:796–808. doi: 10.1016/j.jcmg.2021.12.008
99. Zheng C, Huang L, Luo W, Yu W, Hu X, Guan X, et al. Inhibition of STAT3 in tubular epithelial cells prevents kidney fibrosis and nephropathy in STZ-induced diabetic mice. *Cell Death Dis.* (2019) 10:848–61. doi: 10.1038/s41419-019-2085-0
100. Ritchie RH, Abel ED. Basic mechanisms of diabetic heart disease. *Circ Res.* (2020) 126:1501–25. doi: 10.1161/circresaha.120.315913
101. Zhang Y, Cao Y, Zheng R, Xiong Z, Zhu Z, Gao F, et al. Fibroblast-specific activation of Rnd3 protects against cardiac remodeling in diabetic cardiomyopathy via suppression of Notch and TGF- β signaling. *Theranostics.* (2022) 12:7250–66. doi: 10.7150/thno.77043
102. Nakamura K, Miyoshi T, Yoshida M, Akagi S, Saito Y, Ejiri K, et al. Pathophysiology and treatment of diabetic cardiomyopathy and heart failure in patients with diabetes mellitus. *Int J Mol Sci.* (2022) 23:3587–600. doi: 10.3390/ijms23073587
103. Lafuse WP, Wozniak DJ, Rajaram MVS. Role of cardiac macrophages on cardiac inflammation, fibrosis and tissue repair. *Cells.* (2020) 10:51–77. doi: 10.3390/cells10010051
104. Geng K, Ma X, Jiang Z, Huang W, Gu J, Wang P, et al. High glucose-induced STING activation inhibits diabetic wound healing through promoting M1 polarization of macrophages. *Cell Death Discov.* (2023) 9:136–51. doi: 10.1038/s41420-023-01425-x
105. Zhang S, Zhu X, Chen Y, Wen Z, Shi P, Ni Q. The role and therapeutic potential of macrophages in the pathogenesis of diabetic cardiomyopathy. *Front Immunol.* (2024) 15:1393392–405. doi: 10.3389/fimmu.2024.1393392
106. Nishida K, Otsu K. Inflammation and metabolic cardiomyopathy. *Cardiovasc Res.* (2017) 113:389–98. doi: 10.1093/cvr/cvx012
107. Feng B, Chen S, Gordon AD, Chakrabarti S. miR-146a mediates inflammatory changes and fibrosis in the heart in diabetes. *J Mol Cell Cardiol.* (2017) 105:70–6. doi: 10.1016/j.jmcc.2017.03.002
108. He L, Weber KJ, Schilling JD. Glutamine modulates macrophage lipotoxicity. *Nutrients.* (2016) 8:215–28. doi: 10.3390/nu8040215
109. Zhao Y, Guo Y, Jiang Y, Zhu X, Liu Y, Zhang X. Mitophagy regulates macrophage phenotype in diabetic nephropathy rats. *Biochem Biophys Res Commun.* (2017) 494:42–50. doi: 10.1016/j.bbrc.2017.10.088
110. Mahmoudi A, Firouzjaei AA, Darijani F, Navashenaq JG, Taghizadeh E, Darroudi M, et al. Effect of diabetes on efferocytosis process. *Mol Biol Rep.* (2022) 49:10849–63. doi: 10.1007/s11033-022-07725-2
111. Luo M, Zhao F, Cheng H, Su M, Wang Y. Macrophage polarization: an important role in inflammatory diseases. *Front Immunol.* (2024) 15:1352946–61. doi: 10.3389/fimmu.2024.1352946
112. Tan X, Hu L, Shu Z, Chen L, Li X, Du M, et al. Role of CCR2 in the development of streptozotocin-treated diabetic cardiomyopathy. *Diabetes.* (2019) 68:2063–73. doi: 10.2337/db18-1231
113. González A, Schelbert EB, Diez J, Butler J. Myocardial interstitial fibrosis in heart failure: biological and translational perspectives. *J Am Coll Cardiol.* (2018) 71:1696–706. doi: 10.1016/j.jacc.2018.02.021
114. Gu J, Yin ZF, Zhang HL, Fan YQ, Zhang JF, Wang CQ. Characteristics and outcomes of transitions among heart failure categories: a prospective observational cohort study. *ESC Heart failure.* (2020) 7:616–25. doi: 10.1002/ehf2.12619
115. Špinar J, Špinarová L, Vitovec J. Pathophysiology, causes and epidemiology of chronic heart failure. *Vnitř Lek.* (2018) 64:834–8. doi: 10.36290/vnl.2018.114
116. Santos-Gallego CG, Requena-Ibanez JA, San Antonio R, Ishikawa K, Watanabe S, Picatoste B, et al. Empagliflozin ameliorates adverse left ventricular remodeling in nondiabetic heart failure by enhancing myocardial energetics. *J Am Coll Cardiol.* (2019) 73:1931–44. doi: 10.1016/j.jacc.2019.01.056
117. Aryee EK, Ozkan B, Ndumele CE. Heart failure and obesity: the latest pandemic. *Prog Cardiovasc Dis.* (2023) 78:43–8. doi: 10.1016/j.pcad.2023.05.003
118. Revelo XS, Parthiban P, Chen C, Barrow F, Fredrickson G, Wang H, et al. Cardiac resident macrophages prevent fibrosis and stimulate angiogenesis. *Circ Res.* (2021) 129:1086–101. doi: 10.1161/circresaha.121.319737
119. Hanna A, Frangogiannis NG. Inflammatory cytokines and chemokines as therapeutic targets in heart failure. *Cardiovasc Drugs Ther.* (2020) 34:849–63. doi: 10.1007/s10557-020-07071-0
120. Hu S, Yang M, Huang S, Zhong S, Zhang Q, Ding H, et al. Different roles of resident and non-resident macrophages in cardiac fibrosis. *Front Cardiovasc Med.* (2022) 9:818188–97. doi: 10.3389/fcvm.2022.818188
121. Murphy SP, Kakkar R, McCarthy CP, Januzzi JL Jr. Inflammation in heart failure: JACC state-of-the-art review. *J Am Coll Cardiol.* (2020) 75:1324–40. doi: 10.1016/j.jacc.2020.01.014
122. Lu H, Chen R, Barrie PA, Tian Y, Zhang S, Xu H, et al. Fibroblast transdifferentiation promotes conversion of M1 macrophages and replenishment of cardiac resident macrophages following cardiac injury in mice. *Eur J Immunol.* (2020) 50:795–808. doi: 10.1002/eji.201948414
123. Chaikijurajai T, Tang WHW. Reappraisal of inflammatory biomarkers in heart failure. *Curr Heart Fail Rep.* (2020) 17:9–19. doi: 10.1007/s11897-019-00450-1
124. Abraitte A, Aukrust P, Kou L, Anand IS, Young J, McMurray JJV, et al. T cell and monocyte/macrophage activation markers associate with adverse outcome, but give limited prognostic value in anemic patients with heart failure: results from RED-HF. *Clin Res Cardiol.* (2019) 108:133–41. doi: 10.1007/s00392-018-1331-2
125. Sun F, Yuan L, Wang Z, Cui X, Lv N, Zhang T, et al. Cardiac sympathetic overdrive, M2 macrophage activation and fibroblast heterogeneity are associated with cardiac remodeling in a chronic pressure overload rat model of HFpEF. *Front Pharmacol.* (2024) 15:1364758–71. doi: 10.3389/fphar.2024.1364758
126. Shen JL, Xie XJ. Insight into the pro-inflammatory and profibrotic role of macrophage in heart failure with preserved ejection fraction. *J Cardiovasc Pharmacol.* (2020) 76:276–85. doi: 10.1097/fjc.0000000000000858
127. Yildiz M, Oktay AA, Stewart MH, Milani RV, Ventura HO, Lavie CJ. Left ventricular hypertrophy and hypertension. *Prog Cardiovasc Dis.* (2020) 63:10–21. doi: 10.1016/j.pcad.2019.11.009
128. Kruszewska J, Cudnoch-Jedrzejewska A, Czarzasta K. Remodeling and fibrosis of the cardiac muscle in the course of obesity-pathogenesis and involvement of the extracellular matrix. *Int J Mol Sci.* (2022) 23:4195–223. doi: 10.3390/ijms23084195
129. Nakamura M, Sadoshima J. Mechanisms of physiological and pathological cardiac hypertrophy. *Nat Rev Cardiol.* (2018) 15:387–407. doi: 10.1038/s41569-018-0007-y
130. Ferreira NS, Tostes RC, Paradis P, Schiffrin EL. Aldosterone, inflammation, immune system, and hypertension. *Am J Hypertens.* (2021) 34:15–27. doi: 10.1093/ajh/hpaa137

131. Mikolajczyk TP, Szczepaniak P, Vidler F, Maffia P, Graham GJ, Guzik TJ. Role of inflammatory chemokines in hypertension. *Pharmacol Ther.* (2021) 223:107799–919. doi: 10.1016/j.pharmthera.2020.107799
132. Pan CT, Wu XM, Tsai CH, Chang YY, Chen ZW, Chang CC, et al. Hemodynamic and non-hemodynamic components of cardiac remodeling in primary aldosteronism. *Front Endocrinol (Lausanne).* (2021) 12:646097–108. doi: 10.3389/fendo.2021.646097
133. Yang XF, Wang H, Huang Y, Huang JH, Ren HL, Xu Q, et al. Myeloid angiotensin II type 1 receptor mediates macrophage polarization and promotes vascular injury in DOCA/salt hypertensive mice. *Front Pharmacol.* (2022) 13:879693–705. doi: 10.3389/fphar.2022.879693
134. Cai R, Hao Y, Liu YY, Huang L, Yao Y, Zhou MS. Tumor necrosis factor alpha deficiency improves endothelial function and cardiovascular injury in deoxycorticosterone acetate/salt-hypertensive mice. *BioMed Res Int.* (2020) 2020:3921074–83. doi: 10.1155/2020/3921074
135. Kain D, Amit U, Yagil C, Landa N, Naftali-Shani N, Molotski N, et al. Macrophages dictate the progression and manifestation of hypertensive heart disease. *Int J Cardiol.* (2016) 203:381–95. doi: 10.1016/j.ijcard.2015.10.126
136. Liu YY, Luo J, Cai R, Zhang J, Xu Q, Tian Y, et al. Macrophage depletion improves endothelial insulin resistance and protects against cardiovascular injury in salt-sensitive hypertension. *BioMed Res Int.* (2020) 2020:5073762–72. doi: 10.1155/2020/5073762
137. Huang L, Wang A, Hao Y, Li W, Liu C, Yang Z, et al. Macrophage depletion lowered blood pressure and attenuated hypertensive renal injury and fibrosis. *Front Physiol.* (2018) 9:473–83. doi: 10.3389/fphys.2018.00473
138. Yihui C, Yanfeng G. Inflammatory markers in patients with hypertension. *Br J Hosp Med (Lond).* (2023) 84:1–8. doi: 10.12968/hmed.2022.0531
139. Zhang YL, Bai J, Yu WJ, Lin QY, Li HH. CD11b mediates hypertensive cardiac remodeling by regulating macrophage infiltration and polarization. *J Adv Res.* (2024) 55:17–31. doi: 10.1016/j.jare.2023.02.010
140. Takimoto-Ohnishi E, Murakami K. Renin-angiotensin system research: from molecules to the whole body. *J Physiol Sci.* (2019) 69:581–7. doi: 10.1007/s12576-019-00679-4
141. Wright MD, Binger KJ. Macrophage heterogeneity and renin-angiotensin system disorders. *Pflugers Arch.* (2017) 469:445–54. doi: 10.1007/s00424-017-1940-z
142. Abdolmaleki G, Taheri MA, Paridehpour S, Mohammadi NM, Tabatabaei YA, Mousavi T, et al. A comparison between SARS-CoV-1 and SARS-CoV2: an update on current COVID-19 vaccines. *Daru.* (2022) 30:379–406. doi: 10.1007/s40199-022-00446-8
143. Pesce M, Agostoni P, Bøtker HE, Brundel B, Davidson SM, Caterina R, et al. COVID-19-related cardiac complications from clinical evidences to basic mechanisms: opinion paper of the ESC Working Group on Cellular Biology of the Heart. *Cardiovasc Res.* (2021) 117:2148–60. doi: 10.1093/cvr/cvab201
144. Desai AD, Lavelle M, Boursiquot BC, Wan EY. Long-term complications of COVID-19. *Am J Physiol Cell Physiol.* (2022) 322:C1–11. doi: 10.1152/ajpcell.00375.2021
145. Luszczek ER, Ingraham NE, Karam BS, Proper J, Siegel L, Helgeson ES, et al. Characterizing COVID-19 clinical phenotypes and associated comorbidities and complication profiles. *PLoS One.* (2021) 16:e0248956–72. doi: 10.1371/journal.pone.0248956
146. Ali AM, Rostam HM, Fatah MH, Noori CM, Ali KM, Tawfeeq HM. Serum troponin, D-dimer, and CRP level in severe coronavirus (COVID-19) patients. *Immun Inflamm Dis.* (2022) 10:e582–91. doi: 10.1002/iid3.582
147. Fox SE, Falgout L, Vander Heide RS. COVID-19 myocarditis: quantitative analysis of the inflammatory infiltrate and a proposed mechanism. *Cardiovasc Pathol.* (2021) 54:107361–8. doi: 10.1016/j.carpath.2021.107361
148. Flikweert AW, Kobold ACM, van der Sar-van der Brugge S, Heeringa P, Rodenhuis-Zybert IA, Bijzet J, et al. Circulating adipokine levels and COVID-19 severity in hospitalized patients. *Int J Obes (Lond).* (2023) 47:126–37. doi: 10.1038/s41366-022-01246-5
149. Abou Hassan OK, Sheng CC, Wang TKM, Cremer PC. SARS-CoV-2 myocarditis: insights into incidence, prognosis, and therapeutic implications. *Curr Cardiol Rep.* (2021) 23:129–42. doi: 10.1007/s11886-021-01551-x
150. Sang CJ, Burkett A, Heindl B, Litovsky SH, Prabhu SD, Benson PV, et al. Cardiac pathology in COVID-19: a single center autopsy experience. *Cardiovasc Pathol.* (2021) 54:107370–6. doi: 10.1016/j.carpath.2021.107370
151. Sharifkashani S, Bafrani MA, Khaboushan AS, Pirzadeh M, Kheirandish A, Yavarpour Bali H, et al. Angiotensin-converting enzyme 2 (ACE2) receptor and SARS-CoV-2: Potential therapeutic targeting. *Eur J Pharmacol.* (2020) 884:173455–60. doi: 10.1016/j.ejphar.2020.173455
152. Huynh TV, Rethi L, Lee TW, Higa S, Kao YH, Chen YJ. Spike protein impairs mitochondrial function in human cardiomyocytes: mechanisms underlying cardiac injury in COVID-19. *Cells.* (2023) 12:877–93. doi: 10.3390/cells12060877
153. Yamamoto Y, Inoue T, Inoue M, Murai M, Fukasawa M, Kaneko MK, et al. SARS-CoV-2 spike protein mutation at cysteine-488 impairs its golgi localization and intracellular S1/S2 processing. *Int J Mol Sci.* (2022) 23–25. doi: 10.3390/ijms232415834
154. Vargas RA, Varela Millán JM, Fajardo Bonilla E. Renin-angiotensin system: Basic and clinical aspects-A general perspective. *Endocrinol Diabetes Nutr (Engl Ed).* (2022) 69:52–62. doi: 10.1016/j.endien.2022.01.005
155. Han B, Lv Y, Moser D, Zhou X, Woehrle T, Han L, et al. ACE2-independent SARS-CoV-2 virus entry through cell surface GRP78 on monocytes - evidence from a translational clinical and experimental approach. *EBioMedicine.* (2023) 98:104869–86. doi: 10.1016/j.ebiom.2023.104869
156. Karki R, Kanneganti TD. Innate immunity, cytokine storm, and inflammatory cell death in COVID-19. *J Transl Med.* (2022) 20:542–79. doi: 10.1186/s12967-022-03767-z
157. Aluru JS, Barsouk A, Saginala K, Rawla P, Barsouk A. Valvular heart disease epidemiology. *Med Sci (Basel).* (2022) 10:32–43. doi: 10.3390/medsci10020032
158. Pavlides GS, Chatzizisis YS, Porter TR. Integrating hemodynamics with ventricular and valvular remodeling in aortic stenosis. A paradigm shift in therapeutic decision making. *Am Heart J.* (2022) 254:66–76. doi: 10.1016/j.jahj.2022.08.004
159. Kim AJ, Xu N, Umeyama K, Hulin A, Ponny SR, Vagnozzi RJ, et al. Deficiency of circulating monocytes ameliorates the progression of myxomatous valve degeneration in marfan syndrome. *Circulation.* (2020) 141:132–46. doi: 10.1161/circulationaha.119.042391
160. Yamazato R, Yamamoto H, Tadehara F, Teragawa H, Kurisu S, Dohi Y, et al. Association between aortic valve calcification and myocardial ischemia, especially in asymptomatic patients. *J Nucl Med.* (2012) 53:1216–21. doi: 10.2967/jnumed.111.099275
161. Kim AJ, Xu N, Yutzy KE. Macrophage lineages in heart valve development and disease. *Cardiovasc Res.* (2021) 117:663–73. doi: 10.1093/cvr/cvaa062
162. Lyu T, Liu Y, Li B, Xu R, Guo J, Zhu D. Single-cell transcriptomics reveals cellular heterogeneity and macrophage-to-mesenchymal transition in bicuspid aortic valve disease. *Biol Direct.* (2023) 18:35–53. doi: 10.1186/s13062-023-00390-w
163. Hulin A, Anstine LJ, Kim AJ, Potter SJ, Defalco T, Lincoln J, et al. Macrophage transitions in heart valve development and myxomatous valve disease. *Arterioscler Thromb Vasc Biol.* (2018) 38:636–44. doi: 10.1161/atvbaha.117.310667
164. Xu N, Alfieri CM, Yu Y, Guo M, Yutzy KE. Wnt signaling inhibition prevents postnatal inflammation and disease progression in mouse congenital myxomatous valve disease. *Arterioscler Thromb Vasc Biol.* (2024) 44:1540–54. doi: 10.1161/atvbaha.123.320388
165. Reddy YNV, Borlaug BA, Gersh BJ. Management of atrial fibrillation across the spectrum of heart failure with preserved and reduced ejection fraction. *Circulation.* (2022) 146:339–57. doi: 10.1161/circulationaha.122.057444
166. Bizhanov KA, Baimbetov AK, Sarsenbayeva AB, Lyan E. Atrial fibrillation: Epidemiology, pathophysiology, and clinical complications (literature review). *J Cardiovasc Electrophysiol.* (2023) 34:153–65. doi: 10.1111/jce.15759
167. Rahm AK, Lugenbiel P, Schweizer PA, Katus HA, Thomas D. Role of ion channels in heart failure and channelopathies. *Biophys Rev.* (2018) 10:1097–106. doi: 10.1007/s12551-018-0442-3
168. Murakata Y, Yamagami F, Murakoshi N, Xu D, Song Z, Li S, et al. Electrical, structural, and autonomic atrial remodeling underlies atrial fibrillation in inflammatory atrial cardiomyopathy. *Front Cardiovasc Med.* (2022) 9:1075358–66. doi: 10.3389/fcvm.2022.1075358
169. Wijesurendra RS, Casadei B. Mechanisms of atrial fibrillation. *Heart.* (2019) 105:1860–7. doi: 10.1136/heartjnl-2018-314267
170. Begieneman MP, Emmens RW, Rijvers L, Kubat B, Paulus WJ, Vonk AB, et al. Ventricular myocarditis coincides with atrial myocarditis in patients. *Cardiovasc Pathol.* (2016) 25:141–8. doi: 10.1016/j.carpath.2015.12.001
171. Smorodinova N, Bláha M, Melenovský V, Rozsivalová K, Přidal J, Ďurišová M, et al. Analysis of immune cell populations in atrial myocardium of patients with atrial fibrillation or sinus rhythm. *PLoS One.* (2017) 12:e0172691–702. doi: 10.1371/journal.pone.0172691
172. Hazarapetyan L, Zelveian PH, Grigoryan S. Inflammation and coagulation are two interconnected pathophysiological pathways in atrial fibrillation pathogenesis. *J Inflamm Res.* (2023) 16:4967–75. doi: 10.2147/jir.S429892
173. Imazio M, Brucato A, Ferrazzi P, Rovere ME, Gandino A, Cemin R, et al. Colchicine reduces postoperative atrial fibrillation: results of the Colchicine for the Prevention of the Postpericardiotomy Syndrome (COPPS) atrial fibrillation substudy. *Circulation.* (2011) 124:2290–5. doi: 10.1161/circulationaha.111.026153
174. Krisai P, Blum S, Schnabel RB, Sticherling C, Kühne M, Von Felten S, et al. Canakinumab after electrical cardioversion in patients with persistent atrial fibrillation: A pilot randomized trial. *Circ Arrhythm Electrophysiol.* (2020) 13:e008197–203. doi: 10.1161/circep.119.008197
175. Taufiq F, Li P, Miake J, Hisatome I. Hyperuricemia as a risk factor for atrial fibrillation due to soluble and crystallized uric acid. *Circ Rep.* (2019) 1:469–73. doi: 10.1253/circep.CR-19-0088
176. Rao F, Xue YM, Wei W, Yang H, Liu FZ, Chen SX, et al. Role of tumor necrosis factor- α in the regulation of T-type calcium channel current in HL-1 cells. *Clin Exp Pharmacol Physiol.* (2016) 43:706–11. doi: 10.1111/1440-1681.12585
177. Lyu J, Huang J, Wu J, Yu T, Wei X, Lei Q. Lack of macrophage migration inhibitory factor reduces susceptibility to ventricular arrhythmias during the acute phase of myocardial infarction. *J Inflamm Res.* (2021) 14:1297–311. doi: 10.2147/jir.S304553
178. Diekmann J, Bengel FM. Cardiac applications of fibroblast activation protein imaging. *PET Clin.* (2023) 18:389–96. doi: 10.1016/j.cpet.2023.03.004

179. Gawalko M, Balsam P, Lodziński P, Grabowski M, Krzowski B, Opolski G, et al. Cardiac arrhythmias in autoimmune diseases. *Circ J*. (2020) 84:685–94. doi: 10.1253/circj.CJ-19-0705
180. Lubos N, van der Gaag S, Gerçek M, Kant S, Leube RE, Krusche CA. Inflammation shapes pathogenesis of murine arrhythmogenic cardiomyopathy. *Basic Res Cardiol*. (2020) 115:42–60. doi: 10.1007/s00395-020-0803-5
181. Sager HB, Hulsmans M, Lavine KJ, Moreira MB, Heidt T, Courties G, et al. Proliferation and recruitment contribute to myocardial macrophage expansion in chronic heart failure. *Circ Res*. (2016) 119:853–64. doi: 10.1161/circresaha.116.309001
182. Leid J, Carrelha J, Boukarabila H, Epelman S, Jacobsen SE, Lavine KJ. Primitive embryonic macrophages are required for coronary development and maturation. *Circ Res*. (2016) 118:1498–511. doi: 10.1161/circresaha.115.308270
183. Cho DI, Kim MR, Jeong HY, Jeong HC, Jeong MH, Yoon SH, et al. Mesenchymal stem cells reciprocally regulate the M1/M2 balance in mouse bone marrow-derived macrophages. *Exp Mol Med*. (2014) 46:e70–8. doi: 10.1038/emm.2013.135
184. Gomez D, Baylis RA, Durgin BG, Newman A, Alencar GF, Mahan S, et al. Interleukin-1 β has atheroprotective effects in advanced atherosclerotic lesions of mice. *Nat Med*. (2018) 24:1418–29. doi: 10.1038/s41591-018-0124-5
185. Shintani Y, Ito T, Fields L, Shiraishi M, Ichihara Y, Sato N, et al. IL-4 as a repurposed biological drug for myocardial infarction through augmentation of reparative cardiac macrophages: proof-of-concept data in mice. *Sci Rep*. (2017) 7:6877–90. doi: 10.1038/s41598-017-07328-z
186. Halimani N, Nesterchuk M, Andreichenko IN, Tsitrina AA, Elchaninov A, Lokhonina A, et al. Phenotypic alteration of BMDM *in vitro* using small interfering RNA. *Cells*. (2022) 11:2498–513. doi: 10.3390/cells11162498
187. Le Z, Ramos MC, Shou Y, Li RR, Cheng HS, Jang CJ, et al. Bioactive sucralfate-based microneedles promote wound healing through reprogramming macrophages and protecting endogenous growth factors. *Biomaterials*. (2024) 311–26:122700. doi: 10.1016/j.biomaterials.2024.122700
188. Zhang S, Weinberg S, Deberge M, Gainullina A, Schipma M, Kinchen JM, et al. Efferocytosis fuels requirements of fatty acid oxidation and the electron transport chain to polarize macrophages for tissue repair. *Cell Metab*. (2019) 29:443–56.e445. doi: 10.1016/j.cmet.2018.12.004
189. Mihaila AC, Ciortan L, Tucureanu MM, Simionescu M, Butoi E. Anti-inflammatory neutrophils reprogram macrophages toward a pro-healing phenotype with increased efferocytosis capacity. *Cells*. (2024) 13:208–22. doi: 10.3390/cells13030208
190. Liu Y, Wu M, Zhong C, Xu B, Kang L. M2-like macrophages transplantation protects against the doxorubicin-induced heart failure via mitochondrial transfer. *Biomater Res*. (2022) 26:14–27. doi: 10.1186/s40824-022-00260-y
191. Zhang H, Reilly MP. Human induced pluripotent stem cell-derived macrophages for unraveling human macrophage biology. *Arterioscler Thromb Vasc Biol*. (2017) 37:2000–6. doi: 10.1161/atvbaha.117.309195
192. Xia Y, Rao L, Yao H, Wang Z, Ning P, Chen X. Engineering macrophages for cancer immunotherapy and drug delivery. *Adv Mater*. (2020) 32:e2002054–73. doi: 10.1002/adma.202002054
193. Eilers A, Witt S, Walter J. Aptamer-modified nanoparticles in medical applications. *Adv Biochem Eng Biotechnol*. (2020) 174:161–93. doi: 10.1007/10_2020_124
194. Chen W, Schilperoord M, Cao Y, Shi J, Tabas I, Tao W. Macrophage-targeted nanomedicine for the diagnosis and treatment of atherosclerosis. *Nat Rev Cardiol*. (2022) 19:228–49. doi: 10.1038/s41569-021-00629-x
195. Hu D, Li R, Li Y, Wang M, Wang L, Wang S, et al. Inflammation-targeted nanomedicines alleviate oxidative stress and reprogram macrophages polarization for myocardial infarction treatment. *Adv Sci (Weinh)*. (2024) 11:e2308910–30. doi: 10.1002/advs.202308910
196. Wen JH, Li DY, Liang S, Yang C, Tang JX, Liu HF. Macrophage autophagy in macrophage polarization, chronic inflammation and organ fibrosis. *Front Immunol*. (2022) 13:946832–45. doi: 10.3389/fimmu.2022.946832
197. Higashikuni Y, Liu W, Numata G, Tanaka K, Fukuda D, Tanaka Y, et al. NLRP3 inflammasome activation through heart-brain interaction initiates cardiac inflammation and hypertrophy during pressure overload. *Circulation*. (2023) 147:338–55. doi: 10.1161/circulationaha.122.060860
198. Ridker PM, Everett BM, Thuren T, Macfadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med*. (2017) 377:1119–31. doi: 10.1056/NEJMoa1707914
199. Roubille F, Bouabdallaoui N, Kouz S, Waters DD, Diaz R, Maggioni AP, et al. Low-dose colchicine in patients with type 2 diabetes and recent myocardial infarction from the LoDoCo2 randomized controlled trial (COLCOT). *Diabetes Care*. (2024) 47:467–70. doi: 10.2337/dc23-1825
200. Mohammadnia N, Los J, Opstal TSJ, Fiolet ATL, Eikelboom JW, Mosterd A, et al. Colchicine and diabetes in patients with chronic coronary artery disease: insights from the LoDoCo2 randomized controlled trial. *Front Cardiovasc Med*. (2023) 10:1244529-. doi: 10.3389/fcvm.2023.1244529
201. Broch K, Anstensrud AK, Woxholt S, Sharma K, Tøllefsen IM, Bendz B, et al. Randomized trial of interleukin-6 receptor inhibition in patients with acute ST-segment elevation myocardial infarction. *J Am Coll Cardiol*. (2021) 77:1845–55. doi: 10.1016/j.jacc.2021.02.049
202. Silvis MJM, Demkes EJ, Fiolet ATL, Dekker M, Bosch L, Van Hout GPJ, et al. Immunomodulation of the NLRP3 inflammasome in atherosclerosis, coronary artery disease, and acute myocardial infarction. *J Cardiovasc Trans Res*. (2021) 14:23–34. doi: 10.1007/s12265-020-10049-w
203. Simonen P, Ulander L, Eklund KK, Niemi M, Backman JT, Gylling H, et al. The effect of hydroxychloroquine on cholesterol synthesis depends on the profile of cholesterol metabolism. A controlled clinical study. *Atheroscler Plus*. (2024) 55:93–7. doi: 10.1016/j.athplu.2024.02.002
204. Yap J, Irei J, Lozano-Gerona J, Vanapruks S, Bishop T, Boisvert WA. Macrophages in cardiac remodelling after myocardial infarction. *Nat Rev Cardiol*. (2023) 20:373–85. doi: 10.1038/s41569-022-00823-5
205. Mouton AJ, Aitken NM, Moak SP, Do Carmo JM, Da Silva AA, Omoto ACM, et al. Temporal changes in glucose metabolism reflect polarization in resident and monocyte-derived macrophages after myocardial infarction. *Front Cardiovasc Med*. (2023) 10:1136252–66. doi: 10.3389/fcvm.2023.1136252
206. Zhang FS, He QZ, Qin CH, Little PJ, Weng JP, Xu SW. Therapeutic potential of colchicine in cardiovascular medicine: a pharmacological review. *Acta Pharmacol Sin*. (2022) 43:2173–90. doi: 10.1038/s41401-021-00835-w
207. Hitscherich P, Lee EJ. Crosstalk between cardiac cells and macrophages postmyocardial infarction: insights from *in vitro* studies. *Tissue Eng Part B Rev*. (2021) 27:475–85. doi: 10.1089/ten.TEB.2020.0198
208. Vallejo J, Cochain C, Zernecke A, Ley K. Heterogeneity of immune cells in human atherosclerosis revealed by scRNA-Seq. *Cardiovasc Res*. (2021) 117:2537–43. doi: 10.1093/cvr/cvab260
209. Ko T, Nomura S, Yamada S, Fujita K, Fujita T, Satoh M, et al. Cardiac fibroblasts regulate the development of heart failure via Htra3-TGF- β -IGFBP7 axis. *Nat Commun*. (2022) 13:3275–91. doi: 10.1038/s41467-022-30630-y
210. Perbellini F, Thum T. Living myocardial slices: a novel multicellular model for cardiac translational research. *Eur Heart J*. (2020) 41:2405–8. doi: 10.1093/eurheartj/ehz779